8 Macrovascular Complications in Diabetes

39 The Pathogenesis of Macrovascular Complications Including Atherosclerosis in Diabetes

Riccardo Candido^{1,2}, Mark E. Cooper¹ & Karin A.M. Jandeleit-Dahm¹

¹Baker IDI Heart and Diabetes Institute, Danielle Alberti JDRF Center for Diabetes Complications, Melbourne, Australia ²Diabetic Center, A.S.S. 1 Triestina, Trieste, Italy

Keypoints

- Diabetes accounts for 75–90% of excess coronary artery disease risk seen in people with diabetes and enhances the effects of other cardiovascular risk factors.
- A range of hemodynamic and metabolic factors contribute to macrovascular disease in diabetes.
- The link between glucose control and cardiovascular disease (CVD) is not as strong as that seen with microvascular complications of diabetes.
- There is clear *in vitro* and *in vivo* evidence that glucose exerts direct and indirect toxic effects on the vasculature.
- Glycemic memory describes the deferred long-term injurious effects of prior glycemic status; early glycemic control appears to be important to reduce vascular complications in subsequent decades.
- Specific insulin resistance pathways appear to contribute to atherogenesis in diabetes.
- The accumulation of advanced glycation end-products (AGEs) exerts pro-inflammatory and pro-fibrotic effects on the vasculature via receptor independent and receptor dependent effects. The AGE receptor (RAGE) is pivotally involved in the pathogenesis of diabetes accelerated atherosclerosis.

- The components of the classic renin angiotensin system (RAS) and in particular more recently discovered components such as angiotensin converting enzyme 2 appear to contribute to macrovascular disease in diabetes. Inhibitors of the RAS have consistently demonstrated reduced endothelial dysfunction and atherosclerosis in animal models via suppression of inflammation, fibrosis and oxidative stress. There is also strong clinical evidence for vasculoprotection by RAS blockade.
- Other vasoactive components such as endothelin and urotensin II are also likely to contribute to macrovascular complications in diabetes and interact with the RAS. Furthermore, the role of novel tumor necrosis factor (TNF) related ligands, such as TNF-related apoptosis inducing ligand and osteoprotegerin, and the complement system in atherosclerosis are currently under evaluation.
- Treatments that reduce oxidative stress and inhibit inflammation such as peroxisome proliferator activated receptor agonists have been shown to be anti-atherosclerotic in experimental studies with some evidence, albeit not uniform, also having been obtained in clinical studies.
- A multifactorial approach treating conventional cardiovascular risk factors as well as diabetes specific risk factors is currently viewed as the optimal strategy to reduce the burden of CVD in diabetes.

Epidemiology of diabetic macrovascular complications

Macrovascular complications develop in patients with type 1 (T1DM) [1–3] and type 2 diabetes mellitus (T2DM) [4–6]. This is of a particular concern as the increasing prevalence of diabetes now also affects adolescents and younger adults, thus promoting the earlier development of long-term cardiovascular complications. Even after adjusting for concomitant risk factors such as

hypertension and hyperlipidemia there remains an excess risk for cardiovascular disease (CVD) in people with diabetes [7,8]. Indeed, diabetes itself accounts for 75–90% of the excess coronary artery disease (CAD) risk and enhances the effects of other cardiovascular risk factors. Death from stroke and myocardial infarction (MI) are the leading causes of mortality in T1DM and T2DM [2,9].

A range of hemodynamic and metabolic factors have been considered responsible for the development and progression of macrovascular disease in diabetes (Figure 39.1) [10]. In terms of hemodynamic factors, in particular the hormonal cascade known as the renin angiotensin system (RAS) has been shown to have a pivotal role in diabetes-associated atherosclerosis; however, other vasoactive hormone systems such as the endothelin (ET)

Textbook of Diabetes, 4th edition. Edited by R. Holt, C. Cockram, A. Flyvbjerg and B. Goldstein. © 2010 Blackwell Publishing.

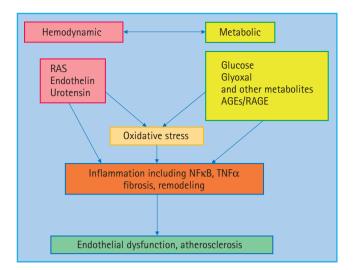


Figure 39.1 Hemodynamic and metabolic mediators contribute to the pathogenesis of diabetic vascular complications. Hemodynamic factors include the renin angiotensin system (RAS), and other vasoactive factors. Metabolic factors include glucose and glucose metabolites such as the formation of advanced glycation end-products (AGEs). Both pathways interact with each other and lead to oxidative stress and inflammation, thus promoting endothelial dysfunction and atherosclerosis. NFkB, nuclear transcription factor kB; TNF- α , tumor necrosis factor α .

[11] and urotensin systems [12,13] have also been implicated in diabetic macrovascular disease. More recently, a role for novel factors such as tumor necrosis factor (TNF) related apoptosis inducing ligand (TRAIL) [14] and the complement system has also been suggested [15].

Pathogenesis of diabetic macrovascular disease

Atherosclerosis is initiated by the adhesion of monocytes to the vascular wall, particularly to endothelial cells, followed by transmigration of monocytes into the subendothelial space [16]. Monocytes differentiate into macrophages which, via uptake of lipids, transform into foam cells and accumulate in the vascular wall. The early atherosclerotic process results in the formation of fatty streak lesions which over time develop into more advanced lesions, characterized by infiltration with vascular smooth muscle cells (VSMC), formation of a necrotic core and further lipid accumulation [16]. In humans, these lesions can demonstrate features of instability and plaque rupture including intraplaque hemorrhages as well as heightened thrombogenicity.

Role of hyperglycemia

A pivotal role for glycemic control and duration of diabetes exposure has been well established for microvascular complications in the UK Prospective Diabetes Study (UKPDS) study [4,17], although the data are not as convincing for macrovascular disease. The importance of glycemic control for macrovascular injury has only recently been extensively investigated in a series of studies that were reported over the last 12 months. It has been suggested that HbA_{1c} acts as an independent and continuous risk factor for macrovascular disease [18–21]; however, this link is not as strong as has been identified with respect to microvascular complications. For an increase in HbA_{1c} from 5.5% (37 mmol/mol) to 9.5% (80 mmol/mol) there is a 10-fold increase in microvascular disease endpoints, whereas the risk for macrovascular disease endpoints increases only twofold.

Clinical trials and hyperglycemia

Recently, clinical trials such as the Action to Control Cardiovascular Risk in Diabetes trial (ACCORD) [22], Action in Diabetes and Vascular disease: PreterAx and DiamicroN-MR Controlled Evaluation (ADVANCE) trial [23] and the Veterans Affairs Diabetes Trial (VADT) [24], which explored tight glycemic control in patients with T2DM with established CVD, have investigated the association between tight glycemic control and cardiovascular endpoints. As observed in the initial reports of the Diabetes Control and Complications Trial (DCCT) in patients with T1DM and in the UKPDS trial in patients with T2DM, tight blood glucose control had little effect on macrovascular outcomes. This was further emphasized by the recent studies including the ADVANCE study [23] and ACCORD trial [22]. Both studies failed to demonstrate significant cardiovascular benefits, with the ACCORD study suggesting possibly deleterious cardiovascular outcomes in association with tight glucose control.

One possible explanation for the lack of a positive effect of tight glycemic control on cardiovascular outcomes may be the short duration of these trials (less than 5 years). Indeed, as best seen with respect to the longer follow-up of the Steno-2 study [25] and a recent long-term follow-up of the UKPDS [26], the benefits of prior intensified cardiovascular risk management including aggressive glucose control may not appear for up to 10 years after initiation of the studies. Thus, in view of the relative lack of an impact on CVD with intensive glucose control, other risk factor modification strategies need to be emphasized.

Direct and indirect glycotoxicity

Hyperglycemia is thought to have direct and indirect toxic effect on vascular cells. It has been suggested that increased glucose levels enter the polyol pathway at an increased flux rate leading to heightened formation of diacylglycerol. In addition, increased flux of glucose into the hexosamine pathway may contribute to glucose mediated vascular injury in diabetes. To investigate specifically hyperglycemia mediated atherosclerosis, aldose reductase, transgenic mice have been investigated [27]. These mice demonstrated increased glucose delivery via the polyol pathway and also showed early atherosclerosis, but did not develop advanced vascular lesions.

In vitro studies

Studies in cell culture experiments have obtained clear evidence that hyperglycemia induces a range of pro-atherogenic effects.

Glucose directly activates monocytes-macrophages in vitro initiating increased expression of cytokines such as interleukin 1ß (IL-1 β) and IL-6 [28]. Furthermore, this leads to protein kinase C (PKC) and nuclear factor kB (NFkB) activation resulting in increased production of reactive oxygen species (ROS). Autooxidation of glucose can also lead to the formation of ROS and can mediate low density lipoprotein (LDL) oxidation. Scavenger receptors on activated macrophages can mediate the uptake of modified lipids such as the pro-atherogenic oxidized LDL. The formation of advanced glycation end-products (AGEs) in the hyperglycemic milieu can lead to the formation of modified albumin which inhibits scavenger receptor class B type 1 mediated efflux of cholesterol to high density lipoprotein (HDL). Therefore, prolonged hyperglycemia can indirectly lead to a range of secondary changes including changes in lipid profile, cellular lipid accumulation and foam cell formation as well as AGE mediated modification of proteins leading to altered cellular structure and function.

Animal models

The study of atherosclerosis in diabetes has long been hampered by the lack of an appropriate animal model. Our group and others have developed a murine model of diabetes associated macrovascular disease, the apoE knockout (KO) mouse rendered diabetic by multiple low doses of streptozotocin injections. This model is currently considered by the National Institutes of Health/Juvenile Diabetes Research Foundation (NIH/JDRF) co-sponsored Animal Models for Diabetes Complications Consortium (AMDCC) to be an appropriate model to study macrovascular disease in diabetes [29].

To address the separate roles of hyperglycemia and dyslipidemia in diabetes-associated atherosclerosis, mice deficient in the LDL receptor were bred with transgenic mice expressing a viral protein under control of the insulin promoter [30]. When infected with the virus, T cells mediate destruction of pancreatic β -cells that express the viral protein, thus closely mimicking the autoimmune response in human T1DM. In these animals, atherosclerosis development was accelerated even on a normal diet suggesting that hyperglycemia was driving atherosclerosis in these mice [30,31]. When animals were placed on a high fat diet, a further acceleration of atherosclerosis occurred suggesting that glucose and lipids may act through synergistic mechanisms to accelerate atherosclerosis. Furthermore, plaque disruption and intraplaque hemorrhages, features of plaque instability and potential plaque rupture were observed in this model [30].

Metabolic memory

The concept of "glycemic memory" describes the deferred effects of prior glycemic status on the subsequent development of diabetic complications. Episodes of poor glycemic control during the earlier stages of diabetes can precipitate or accelerate the development of complications later in the course of diabetes even when glycemic control is subsequently improved. Two studies have provided evidence for such a metabolic imprint, the DCCT/EDIC trial in T1DM [2] and the UKPDS in T2DM [26]. Both studies have shown that poor glycemic control was associated with an increased subsequent burden of complications. Another study in patients with T2DM, the VADT study, compared intensive with standard glucose control (6.9 vs. 8.4%, 52 vs 68 mmol/mol), but did not demonstrate significant cardiovascular protection after patients had been exposed to hyperglycemia for long periods of time (12–15 years) [24]. Therefore, it appears that early glycemic control is crucial for a long-term beneficial outcome on diabetic complications.

The concept of "metabolic memory" raises two important issues. First, hyperglycemia may expose patients to the harmful effects of hyperglycemia years before T2DM is diagnosed. Indeed, 25% of patients show complications at the time of diagnosis of T2DM. Therefore, it has been postulated that early diagnosis and strict glycemic control may be pivotal to reduce the induction of this metabolic memory with subsequent development of long-term diabetic vascular complications. Secondly, glycemic oscillations including peaks and troughs are not reflected in the HbA_{1c} levels [32,33] but may have a key role in mediating growth factor and cytokine expression as well as inducing chromatin remodeling [34].

There are many cellular and molecular processes that may contribute to the mechanisms underlying metabolic memory with most of those relating to glycotoxicity. These pathways include the formation of AGEs, glycation of DNA, increased flux of glucose metabolism, leading to increased oxidative damage, overproduction of PKC- β as well as mitochondrial stress. Recently, it has been shown that transient hyperglycemia induces long-lasting activation of epigenetic changes in the promoter region of the NF κ B subunit p65 in aortic endothelial cells both *in vitro* and *in vivo*. These hyperglycemia induced epigenetic changes and increased p65 expression were prevented by reducing mitochondrial superoxide production or superoxide induced generation of α -oxoaldehydes such as methylglyoxal (Figure 39.2) [34]. These studies suggest that transient hyperglycemia

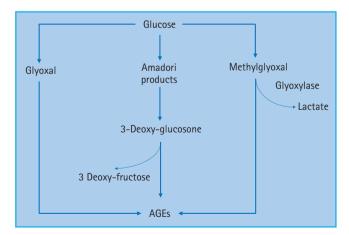


Figure 39.2 The formation of advanced glycation end-products (AGEs) can involve early glucose metabolites such as glyoxal and methylglyoxal, highly reactive dicarbonyls and key precursors of AGEs.

causes persistent atherogenic effects during subsequent normoglycemia by inducing long-lasting changes in chromatin remodeling. Specifically, hyperglycemia results in recruitment of the histone methyl transferase Set7 with increased H3K4 monomethylation of the proximal promoter region of the NFKB subunit p65 gene leading to increased expression of pro-atherogenic pathways including monocyte chemotactic protein 1 (MCP-1) and vascular cell adhesion molecule 1 (VCAM-1) [34]. Subsequent studies in mice, which were initially diabetic but returned spontaneously to normoglycemia, have also demonstrated persistent upregulation of pro-inflammatory genes such as p65 and MCP-1 as a result of prior hyperglycemia. Furthermore, more detailed epigenetic studies revealed that changes in histone modifications as a result of prior hyperglycemia also included changes in H3K9 methylation of the p65 promoter as well as effects on various histone methyl transferases and interestingly also demethylases such as LSD-1 [35].

Insulin resistance

Insulin resistance occurs in T2DM and in patients with impaired glucose tolerance, and has been associated with increased cardio-vascular risk [36–38]. There is now increasing evidence that insulin resistance promotes atherogenesis as an independent risk factor [39–41]. Furthermore, insulin resistance is often associated with a pro-atherogenic lipid profile that includes a high very low-density lipoprotein (VLDL) component, a low HDL and small dense LDL.

In vitro studies have shown that insulin exerts both proatherogenic and anti-atherogenic effects [42,43]. This has led to the hypothesis of pathway specific insulin resistance (Figure 39.3). It has been suggested that insulin resistance towards glucose transport also affects resistance to the antiproliferative effects of insulin, whereas the signaling pathways leading to cellular proliferation remain intact (Figure 39.3). Insulin signaling via phosphatidylinositol 3 kinase (PI3K) has been associated with antiproliferative and antithrombogenic effects with decreased adhesion molecule expression such as MCP-1, VCAM-

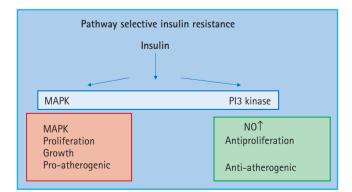


Figure 39.3 Pathway selective insulin resistance leading to endothelial dysfunction and atherosclerosis. MAPK, mitogen activated protein kinase; NO, nitric oxide; PI3 kinase, phosphatidylinositol 3 kinase.

1 and intercellular adhesion molecule 1 (ICAM-1). Furthermore, nitric oxide (NO) production is mediated via the PI3K signal transduction pathway and therefore is impaired in insulin resistance. NO has been shown to reduce LDL oxidation and proliferation of VSMC. In contrast, effects on VSMC proliferation and migration are mediated via the Ras/Raf/MEKK/MAPK signal transduction pathway which is further stimulated in the context of hyperinsulinemia, thus suggesting that pathway specific insulin resistance contributes to the pro-atherogenic effects of insulin. More recently, it has been shown that IL-6 decreases insulin stimulated NO production from endothelial cells via decreased activity of insulin signaling mediated by enhanced TNF-a production [44]. Paradoxically, IL-6 increases insulin stimulated glucose uptake into skeletal muscle and adipose tissue via enhanced insulin signaling, yet IL-6 and insulin have not been linked to increased TNF- α expression in skeletal muscle [44].

Formation of AGE

AGE formation originates from early glycation products, the Schiff bases, which form the more stable Amadori products such as 1-amino-1-deoxyfructose derivatives. The Amadori products undergo further enzymatic modifications resulting in a number of reactive intermediates such as 3-deoxyglucosone and methylglyoxal (Figure 39.2). Methylglyoxal reacts with amino, sulfudryl and guanidine functional groups in proteins causing the browning, denaturation and redox active di-amine cross-linking between lysine residues of the target amino acids. Methylglyoxal also generates hydroimidazolones, N-E-(carboxyethyl) lysine, a homolog of carboxymethyl lysine (CML) and methylglyoxal lysine dimer [45,46]. The AGE-based cross-links are resistant to enzymatic degradation and therefore very stable [47]. The rate of AGE formation is dependent on multiple factors including the ambient concentrations of various sugars including glucose, the extent of oxidative stress and the duration of exposure to these various stimuli [48,49].

Direct effects of vascular AGE accumulation

There is evidence from experimental studies that AGEs directly influence endothelial function [50] as well as enhancing the evolution of macrovascular disease [51,52]. AGEs mediate their effects both directly and via receptor-mediated mechanisms. AGE accumulation in the vascular wall is associated with changes in the structural integrity of proteins, disturbance of their cellular function and degradation of these proteins [28]. AGEs accumulate on many proteins including collagen, albumin and apolipoproteins. Furthermore, the cross-linking of AGEs with matrix molecules can disrupt matrix-matrix and matrix-cell interactions. It has been shown that AGE cross-linking to collagens decreases vascular elasticity and reduces vascular compliance resulting in increased vascular stiffness [53,54]. In addition, AGEs are also able to quench NO [50,55] and generate ROS by stimulating nicotinic acid adenine dinucleotide phosphate (NADPH) oxidase activity [56].

AGE binding proteins

The receptor-mediated effects of AGEs occur via binding to proteins such as RAGE [57], AGE-R1 (p60), AGE-R2 (p90) and AGE-R3 (galectin-3), the ezrin-radixin-moesin (ERM) family of proteins [58] macrophage scavenger receptor ScR-11 and CD-36 [59]. The exact roles of AGE-R1, AGE-R2 and AGE-R3 have not been fully elucidated. Finally, the interaction between AGEs and several macrophage scavenger receptors, such as CD36 [59], has been postulated to also promote atherosclerosis with studies using CD36-knockout mice supporting the view that CD36 promotes atherosclerosis [60].

Receptor for AGEs, RAGE

RAGE is a multiligand signal transduction receptor of the immunoglobulin superfamily of cell surface molecules that acts as a pattern recognition receptor [57]. In addition to binding ligands actively participating in inflammation and immune responses, RAGE serves as an endothelial adhesion receptor for leukocyte integrins and promotes leukocyte recruitment and extravasation of infiltrating cells. Of direct relevance to diabetic macrovascular complications, RAGE is found on endothelial cells and monocytes–macrophages [52,61,62], with RAGE having been implicated in inflammatory lesions in many disorders [63].

Downstream effects of RAGE activation

Engagement of RAGE leads to activation of the pro-inflammatory transcription factor NF κ B (Figure 39.4) [64]. AGE binding to RAGE activates various signaling pathways, including NADPH oxidase [56], MAPKs, p21^{ras} [65], extracellular signal-regulated kinases (ERKs) [66] and PKC causing activation and translocation of NF κ B [64]. Furthermore, expression of RAGE itself can be induced by NF κ B [67]. RAGE expression within tissues is markedly enhanced in response to metabolic disturbances such as diabetes, dyslipidemia, uremia and aging, possibly because of accumulation of AGEs in these conditions [61,63]. In plaques from diabetic apoE KO mice, upregulation of connective tissue growth factor (CTGF) has been demonstrated that appears to be AGE dependent [51] and may be mediated via RAGE. Intracellular accumulation of AGEs may also promote phenotypic conversion of VSMCs into foam cells within atherosclerotic plaques [68].

Studies reducing vascular AGE accumulation

A variety of pharmacologic interventions have been used to reduce the accumulation of AGEs via decreasing the total AGE load or via chemical modification of existing AGEs into inactive forms [69,70]. Aminoguanidine is a potent inhibitor of the formation of AGEs, scavenges reactive dicarbonyl AGE precursors [69,70] and decreases oxidative damage to mitochondrial proteins [71]. In experimental and clinical diabetes, aminoguanidine treatment has been shown to reduce microvascular [72] and more recently macrovascular complications in a model of accelerated atherosclerosis, the diabetic apoE KO mouse [51].

Another potential agent which inhibits AGE accumulation is ALT-711 or alagebrium [73]. Based initially on a range of *in vitro*

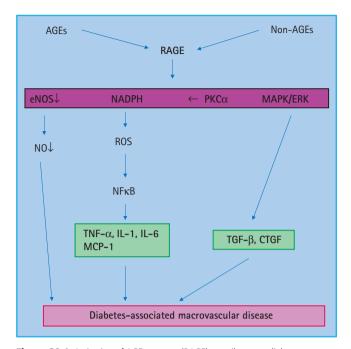


Figure 39.4 Activation of AGE receptor (RAGE) contributes to diabetes associated macrovascular disease via increased production of reactive oxygen species (ROS), decrease in nitric oxide (NO) availability, activation of the nuclear transcription factor κ B (NF κ B) and tumor necrosis factor α (TNF- α) as well as activation of profibrotic growth factors such as transforming growth factor β (TGF- β) and connective tissue growth factor (CTGF) associated with vascular remodeling. eNOS, endothelial nitric oxide synthase; ERK, extracellular signal-regulated kinase; MAPK, mitogen activated protein kinase; MCP, monocyte chemotactic protein; NADPH, nicotinic acid adenine dinucleotide phosphate; PKC, protein kinase C.

studies, this thiazolium compound and its original prototype, phenylthiazolium bromide [74], have been shown to cleave preformed AGEs and thus one of the postulated mechanisms of ALT-711 is as an AGE cross-link breaker [73].

The removal of established cross-links in diabetic rats with ALT-711 has been shown to be associated with reversal of the diabetes-induced increase in large artery stiffness, increased collagen solubility and reduced vascular and cardiac AGE accumulation [54,75-77]. In addition, alagebrium prevents the progression of nephropathy [78,79] possibly via direct inhibition of PKC- α phosphorylation [80], thus reducing renal expression of vascular endothelial growth factor (VEGF). Treatment with ALT 711 in diabetic apoE KO mice was associated with a significant reduction in atherosclerosis [51]. This antiatherosclerotic effect was associated with reduced vascular AGE accumulation and RAGE expression. Alagebrium treatment was also associated with less inflammation and reduced expression of pro-fibrotic growth factors, in particular CTGF [81,82]. In the clinical setting, ALT 711 has been shown to reduce pulse pressure and improve vascular compliance in patients with systolic hypertension [54].

Soluble RAGE

There are three major splice variants of RAGE [83]. First, there is the full-length RAGE receptor; secondly, the N-terminal variant, that does not contain the AGE-binding domain; and, thirdly, a C-terminal splice variant, soluble RAGE (sRAGE), which does not contain the *trans*-membrane and effector domains. It remains controversial as to whether the effects of sRAGE are primarily as a decoy to ligands such as AGEs or whether sRAGE acts as a competitive antagonist to the full-length biologically active RAGE [83,84].

Soluble RAGE and diabetes associated atherosclerosis

Soluble RAGE (sRAGE) has been identified as having therapeutic value in a model of diabetes associated atherosclerosis, the streptozotocin diabetic Apo E KO mouse [85,86]. In the original study, Park et al. [85] reported that diabetic apoE KO mice treated with sRAGE showed a dose-dependent suppression of atherosclerosis and reduced plaque complexity. These beneficial effects were independent of effects on glucose or lipid levels. Furthermore, AGE levels in these diabetic mice were suppressed in a dosedependent manner by sRAGE to levels similar to those seen in non-diabetic animals. Furthermore, the effect of RAGE blockade with sRAGE was investigated in established atherosclerosis [86]. Administration of sRAGE decreased the expression of RAGE, as well as reducing the number of infiltrating inflammatory cells and gene expression for transforming growth factor β (TGF- β), fibronectin and type IV collagen in both the aorta and the kidney in association with reduced plaque area. Therefore, it was concluded by these investigators that RAGE activation not only contributes to lesion formation, but also to the progression of atherosclerosis.

Soluble RAGE treatment has also been reported to be effective in reducing vascular complications in other models of atherosclerosis and diabetes. Atherosclerosis in the LDL receptor -/- mouse made diabetic by streptozotocin injection [87] was significantly attenuated by sRAGE treatment. ApoE KO mice bred onto a *db/db* background, a model of T2DM and deficient leptin receptor signaling, showed increased atherosclerosis which was significantly attenuated by daily sRAGE treatment [88].

To understand better the specific role of RAGE in the genesis of vascular lesions, animals selectively deficient in RAGE/apoE (RAGE –/–) have been created [89,90]. These mice completely lack not only tissue bound full-length RAGE, but also soluble RAGE. As had been predicted by the pharmacologic intervention studies directed towards the RAGE ligands and AGEs, RAGEknockout mice bred onto an apoE –/– background showed a marked reduction in plaque area in the presence and absence of diabetes [89,90]. More recently, the effect of RAGE deletion on atherosclerosis development has been investigated in streptozotocin diabetic RAGE/apoE KO mice and showed a significant reduction in atherosclerotic plaque area when compared with diabetic apoE KO mice expressing RAGE. These vascular changes seen in the streptozotocin diabetic double RAGE/apoE KO mice were associated with reduced inflammation, less accumulation of RAGE ligands such as S100/CML, decreased infiltration by macrophages and T lymphocytes and a reduction in expression of pro-fibrotic and pro-inflammatory growth factors and cytokines [90].

These promising results with RAGE antagonism have encouraged the development of RAGE neutralizing compounds for clinical use. The RAGE modulating agent TTP488 is currently being considered in phase II clinical trials in patients with Alzheimer disease and in patients with diabetic nephropathy. Another compound, TP4000, will soon enter phase I clinical trials (www.Clinicaltrials.gov and www.ttpharma.com); however, currently, the major focus of antagonizing RAGE does not appear to involve studies addressing CVD in the absence or presence of diabetes.

Interaction with the renin angiotensin system

Because diabetic complications appear to be multifactorial in origin and involve interactions between hemodynamic pathways such as the RAS and metabolic pathways such as hyperglycemia and the formation of AGEs, there has been increasing investigation of the potential links between these various pathways (Figure 39.1) [91]. There is increasing evidence that AGE accumulation can induce an upregulation of certain components of the RAS, although these studies have been performed predominantly in the renal context [92]. Furthermore, angiotensin-converting enzyme (ACE) inhibition has been reported to confer its end-organ protective effect, partly via a reduction in AGEs and an increase in sRAGE [93]. Therefore, the status of the RAS could represent a key modulator in AGE-induced diseases. More recently, other therapeutic interventions shown to be anti-atherosclerotic have been reported to exert part of their vasculoprotective effect via inhibition of the AGE/RAGE pathway. These include angiotensin II receptor blockers (ARBs), peroxisome proliferator-activated receptor α (PPAR- α) and PPAR- γ agonists and statins [94–96].

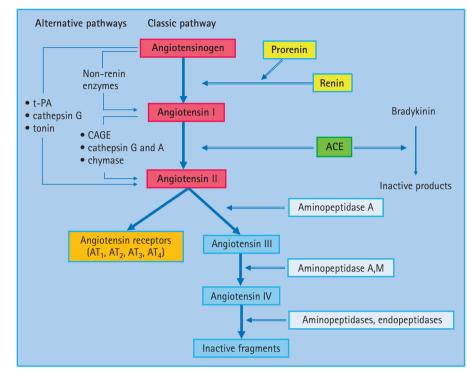
Role of vasoactive hormones in diabetes-related atherosclerosis

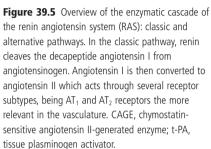
Classic RAS

Renin was described more than 100 years ago by Tigerstedt & Bergman [97]. Nevertheless, our understanding of the RAS is still not complete and has grown increasingly complex over the last decade. The classic pathway is now very well characterized (Figure 39.5). The generation of angiotensin (AT) I and II is not restricted to the systemic circulation but this production also takes place in vascular and other tissues (Figure 39.5) [98].

Novel aspects of the RAS: ACE2

In 2000, a further enzyme associated with the generation of AT peptides was identified – ACE2, a carboxypeptidase with sequence similarity to ACE [99]. ACE2 does not generate AT II but increases the formation of AT 1-7 (Figure 39.6). This heptapeptide causes vasodilatation and has growth inhibitory effects [100]. Further





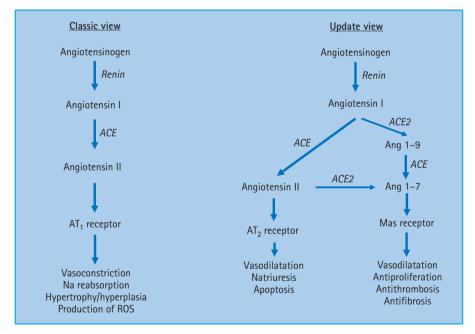


Figure 39.6 New aspects of the renin angiotensin system (RAS) components and its interactions. The classic RAS illustrates the main pathway for angiotensin II (AT II) generation from angiotensin I (AT I) via ACE, with effects being mediated via AT₁ receptor. The updated view illustrates the new components of the RAS in which ACE2 has a role to degrade AT I to AT 1-9, and AT II to the vasodilatator AT 1-7 which acts through the Mas receptor.

research, including into specific inhibitors, is needed to understand the many functions of ACE2 inside and outside the RAS.

AT receptors

The effects of all AT peptides are mediated through specific cell surface receptors (Figures 39.5 and 39.6). The AT_1 receptor mediates most of the effects usually associated with AT II. The

role of the AT_2 receptor subtype remains controversial, but it appears to antagonise certain effects of AT II mediated via the AT_1 receptor [101].

Role of the RAS in macrovascular disease

It is now recognized that local RAS activation has an important role in the pathogenesis of diabetes-induced endothelial

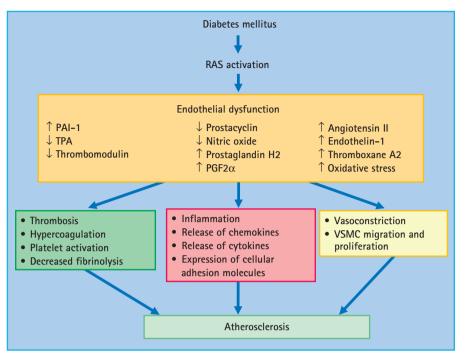


Figure 39.7 Effects of diabetes-induced renin angiotensin system (RAS) activation on mechanisms associated with endothelial dysfunction and atherosclerosis. In diabetes, activation of RAS induces endothelial dysfunction which is characterized by vasoconstriction, inflammation, cellular growth and thrombosis. By losing its protective properties, dysfunctional endothelium is a major promoter of atherogenesis and, consequently, cardiovascular events. PAI-1, plasminogen activator inhibitor-1; TPA, tissue plasminogen activator; VSMC, vascular smooth muscle cells.

dysfunction and atherosclerosis [102–104]. As a result, there is a strong rationale for blockade of the RAS to prevent cardiovascular events in patients with diabetes. In addition, several recent clinical trials [105–110] have suggested that blockade of the RAS may protect against the development of T2DM in at-risk patients.

RAS activation and endothelial dysfunction

Endothelial dysfunction has been considered as an imbalance between vasoconstrictors and vasodilatators in the vascular tone [111] associated with a decrease in NO activity and increased AT II and endothelin (ET) activity [112]. Endothelial dysfunction is characterized by defective endothelium-dependent vasorelaxation in association with inflammatory cytokine-induced increased interactions with blood leukocytes, in which adhesion molecules such as VCAM-1 and chemo-attractants such as MCP-1 are essential [113]. Endothelial dysfunction is strongly associated with atherosclerosis (Figure 39.7) [114,115].

Crucial to the pathogenesis of diabetes-induced endothelial dysfunction in the early stages, in the absence of structural changes of the vessel wall, is the activation of the RAS. Although measurements of components of the RAS in plasma have, in general, suggested suppression of this system in diabetes, there is increasing evidence for activation of the local cardiovascular RAS in the diabetic context [116,117]. It has been demonstrated that there is an increase in ACE expression and activity, in AT II expression, and in AT₁ receptor expression in the aortic wall of diabetic apoE KO mice suggesting a key role for RAS activation in the pathogenesis of diabetes associated endothelial dysfunction [102,118].

RAS and regenerative endothelial cell repair

A role for the RAS has also been demonstrated in affecting the number of regenerative endothelial progenitor cells in patients with diabetes. An increased concentration of circulating endothelial progenitor cells, which are believed to maintain the integrity of the vascular endothelium, has been associated with a favorable cardiovascular outcome in patients with CAD [119]. A study in patients with T2DM [120] suggested that treatment with an ARB (olmesartan) increases the number of regenerative endothelial progenitor cells, which could contribute to the beneficial cardiovascular effects seen with AT₁ receptor blockade.

Role of AT 1-7 and ACE2 in endothelial dysfunction

It has been hypothesized that disruption of the ACE-ACE2 balance may result in abnormal blood pressure, with increased ACE2 expression protecting against hypertension, and ACE2 deficiency causing hypertension (Figure 39.6) [121]. As reported above, it is well established that AT II produces endothelial dysfunction through different pathways, such as increasing oxidative stress and exerting proliferative and pro-thrombotic activities [122]. Recently, it has been observed that AT 1-7, which is generated by the enzyme ACE2, promotes the release of NO and prostaglandins [123,124] and potentiates bradykinin effects in different experimental models [124,125]. In addition, AT 1-7 inhibits growth of VSMC [126], platelet aggregation and thrombosis [127], inflammation, fibrosis [128] and oxidative stress [129] which in turn might lead to restoration of endothelial function. It appears that AT 1-7 can antagonize AT II effects, not only by the stimulation of other vasodilatators but also through AT₁ receptor inhibition. In this regard, Kostenis et al. [130] reported

that the Mas receptor, which is increasingly considered to be the major receptor conferring the biologic effects of AT 1-7, seems to act as a physiologic antagonist of AT_1 receptor and thus counteracting many of the actions of AT II at the endothelial level.

RAS activation and atherosclerosis

Clinical and experimental evidence clearly indicates that activation of the RAS is central to almost all these pro-atherosclerotic pathways (Figure 39.7) [103,131]. Diet et al. [104] observed increased ACE protein accumulation within the atherosclerotic plaque in human coronary arteries suggesting that ACE may contribute to an increased production of local AT II which may participate in the pathophysiology of artery disease. In addition, there is evidence that AT II and the AT₁ receptor are overexpressed in atherosclerotic plaques [102,132,133]. Indeed, blockade of the RAS with an ACE inhibitor or an AT1 receptor antagonist prevents atherosclerosis by mechanisms involving inhibition of pro-inflammatory molecules such as VCAM-1 and MCP-1 and pro-sclerotic and pro-proliferative cytokines such as CTGF and platelet derived growth factor. These observations confirm a key role for RAS activation in the development and progression of atherosclerosis.

Production of pro-inflammatory cytokines, such as IL-1β, TNF-α and IL-6, have a major role in the pathogenesis of atherosclerosis [134]. IL-6, ACE and AT₁ receptors have been detected in stable and unstable atherosclerotic plaques [132,135]. AT II stimulates the redox sensitive nuclear transcription factor, NFκB, which could serve as a unifying signaling system for inflammatory stimuli in atherogenesis through enhanced expression of adhesion molecules ICAM-1 and VCAM-1, E-selectin, MCP-1 and IL-8. Several clinical and experimental studies have demonstrated that both ACE inhibitors and AT₁ receptor blockers decrease the expression of several adhesion molecules, thus confirming that the chronic inflammatory response associated with atherosclerosis appears to be modulated by AT II at every level and can be targeted therapeutically by RAS inhibition [103,116,136].

Moreover, the sustained pro-inflammatory state seems to play an important part in the transformation of a stable atherosclerotic plaque into a vulnerable plaque prone to rupture. Plaque rupture has been connected with activation of matrix metalloproteinases in the fibrous cap of the atherosclerotic lesion [137], and there is evidence that AT II is implicated in matrix metalloproteinase activation, both through a direct action and through induction of pro-inflammatory cytokines such as IL-6.

ACE2 and diabetes accelerated atherosclerosis

Although studies on the expression and activity of ACE2 in atherosclerosis are limited, there is increasing evidence that such more recently identified components of the RAS may be involved in the development and progression of atherosclerosis. Recently, Zulli *et al.* [138] have shown very high expression of ACE2 in endothelial cells, macrophages and α -smooth muscle cells within atherosclerotic plaques in a rabbit model of atherosclerosis. It is not clear whether this increase in ACE2 is in response to injury in an attempt to protect the vessel by increasing levels of AT 1-7.

In the human context, there is evidence for a significant activation of the cardiac RAS after coronary artery occlusion, and it is well known that RAS blockade reduces remodeling and improves survival in humans after an MI. In recent studies [139], cardiac ACE2 expression and activity are also increased with experimental MI. All these observations suggest that an imbalance in the RAS activity has a central role in the pathogenesis of atherosclerosis. Nevertheless, as yet, it is unknown if deficiency of ACE2 contributes to the accelerated atherosclerotic disease process seen in diabetes.

RAS and oxidative stress

There is increasing evidence that the local production of ROS has a pivotal role in atherosclerosis, specifically in the diabetic milieu. Increased vascular superoxide production in aortas from diabetic atherosclerotic apoE KO mice has been demonstrated [94]. These changes were mediated by increased NAD(P)H oxidase (Nox) activity in the aorta with increased expression of various Nox subunits including p47phox, gp91phox and rac-1 [94]. Evidence of increased local vascular ROS generation in diabetes is further supported by the demonstration of increased nitrotyrosine staining in these diabetic plaques. Indeed, interventions that reduce vascular superoxide production such as PPAR- α and PPAR- γ agonists have been associated with reduced plaque formation, further emphasizing the link between vascular oxidative stress and atherosclerosis [94,140]. Furthermore, the deletion of antioxidant enzymes such as glutathione peroxidase, specifically the Gpx1 isoform in the vascular wall, results in an increase in plaque area particularly in the diabetic context via increases in inflammatory mediators including adhesion molecules and chemokines [141]. More recently, it has been shown that treatment of GPx1 deficient mice with the Gpx1 analog, ebselen, reduced oxidative stress variables and atherosclerosis in diabetic GPx1/apoE KO mice [142]. Furthermore, in the Gpx1/apo E double KO mice, there was associated upregulation of RAGE further linking vascular RAGE expression to increased oxidative stress and accelerated atherosclerosis in settings such as diabetes [141].

Therapeutic implications

RAS blockade and endothelial dysfunction

As a result of previous observations, RAS blockade has emerged as an obvious and attractive therapeutic target. Early evidence for a clinical beneficial effect of inhibition of this system on impaired endothelial function was derived from the Trial on Reversing Endothelial Dysfunction (TREND) [143] which showed that ACE inhibition improves endothelial function in subjects with coronary artery disease. Moreover, O'Driscoll *et al.* [144] observed that ACE inhibition with enalapril improved both basal and stimulated NO-dependent endothelial function in normotensive patients with T2DM.

Data are also available for the role of ARBs in improving endothelial function such as the AT₁ receptor antagonist, losartan

[145,146]. Furthermore, in patients with T1DM treated with the ACE inhibitor ramipril or the ARB losartan for 3 weeks [147] improved endothelial dysfunction potentially mediated by increased bradykinin levels with ACE inhibition or increased levels of AT 1-7 levels.

RAS inhibition and cardiovascular protection

There is accumulative evidence that pharmacologic therapy that interrupts the RAS may afford special benefits in reducing CVD in people with diabetes [109,148,149]. In a post hoc subgroup analysis of the Captopril Prevention Project (CAPP) study, the patients with diabetes treated with captopril fared significantly better compared with those treated with conventional therapy (beta-blockers and diuretics) in terms of primary endpoint as well as for MI, all cardiac events and total mortality [148]. These relative beneficial effects of ACE inhibitor therapy were particularly striking in those at highest risk, specifically those with the highest median fasting glucose or those with more elevated blood pressure (BP). This is in contrast to the UKPDS [150], in which there was comparable CVD benefits for patients with T2DM who were randomized to captopril or atenolol, perhaps reflecting a lower CVD risk in the newly diagnosed patients with diabetes studied in the UKPDS.

In a sub-study of the Hypertensive Old People in Edinburgh (HOPE) study, the MICRO-HOPE [151], of the relative risk reduction in the 3577 patients who had diabetes and one other CVD risk factor, there was a risk reduction of 25% for combined CVD events, 37% for CVD mortality, 22% for MI and 33% for stroke. In the Fosinopril versus Amlodipine Cardiovascular Events Trial (FACET) study, the incidence of CVD events was less in hypertensive patients with T2DM treated with fosinopril than the amlodipine-treated group [152]. The European Trial on Reduction of Cardiac Events with Perindopril in Stable Coronary Artery Disease (EUROPA) [153] has shown that ACE inhibition reduced cardiovascular mortality and morbidity in patients with established CAD without left ventricular dysfunction. The data from these trials were pooled with those of the Quinapril Ischemic Event Trial (QUIET) [154] study in a meta-analysis that included a total of 31555 patients [155]. This analysis showed that, compared with placebo, ACE inhibitor therapy produced a significant 14% reduction in all-cause mortality and MI, a 23% reduction in stroke and a 7% statistically significant reduction in revascularization procedures. Recently, the ADVANCE study [23] has shown that the administration of the ACE inhibitor perindopril and the diuretic indapamide in high-risk patients with T2DM induced a reduction in macrovascular outcome when compared with placebo.

Recent reports of trials with ARBs indicate that these agents have CVD protection effects in patients with T2DM similar to those observed with ACE inhibitors. In the Losartan Intervention For Endpoint reduction in hypertension (LIFE) study [109], treatment with losartan in patients with T2DM and left ventricular hypertrophy (LVH) resulted in a significant reduction in death from CVD and in particular all cause mortality. An analysis from the large Reduction of Endpoints in NIDDM with Angiotensin II Antagonist Losartan (RENAAL) study has shown that treatment with losartan in patients with T2DM, nephropathy and LVH reduced the cardiovascular risk to levels similar to those observed in patients without LVH [156]. Recently, the Ongoing Telmisartan Alone and in combination with Ramipril Global Endpoint Trial (ONTARGET) has shown that the ARB telmisartan provides a benefit similar to that of a proven ACE inhibitor such as ramipril in high risk patients with CVD or subjects with diabetes who have end-organ damage. This is a population similar to that examined previously in the HOPE study [157]. Furthermore, in the ONTARGET study the combination of ramipril with telmisartan, despite the further lowering of BP, did not reduce the risk of cardiovascular events, compared with an ACE inhibitor alone, but was associated with additional adverse effects including hypotension and renal dysfunction. Thus, an ACE inhibitor, or possibly an ARB, is an initial antihypertensive agent of choice in patients with diabetes because these agents have been shown to have the capacity to prevent or retard the development of diabetic macrovascular complications, thus significantly reducing cardiovascular mortality and morbidity.

The endothelin system

Endothelin (ET) was first discovered in 1988 by Yanagisawa *et al.* [158] and is one of the most potent vasoconstrictors known. There are three distinct ET genes that encode different mature ET sequences, designated ET-1, ET-2 and ET-3. Big ET-1 is converted into mature 21 amino acid ET-1 by ET-converting enzyme. ET-1 is predominantly present in endothelial cells. In 1990, the ET_A and ET_B receptor subtypes [159], were cloned. ET_A receptors are found in VSMCs and mediate vasoconstriction and cell proliferation (Figure 39.8). ET_B receptors are found in endothelial cells (ET_{B1}) where they mediate vasocilation via the release of NO and on smooth muscle cells, where they may elicit vessel contraction and cell proliferation (Figure 39.8).

Role of ET in diabetic macrovascular complications

Plasma concentrations of ET-1 are increased in patients with T2DM complicated with atherosclerosis, compared with patients without diabetes with atherosclerosis and healthy subjects [160]. Kalogeropoulou *et al.* [161] demonstrated increased ET-1 levels in subjects with diabetes and carotid atherosclerosis. ET stimulates the production and release of inflammatory cytokines from monocytes [162] and enhances the uptake of LDL cholesterol by these cells, promoting a phenotypic change into foam cells [163]. Cytokines released from monocytes–macrophages, in turn, stimulate ET-1 production, providing positive feedback for further cytokine production [164].

In several animal models, both ET_A receptor-selective and nonselective ET_A/ET_B receptor blockade have been shown to inhibit the development of atherosclerotic lesions suggesting that elevated vascular ET-1 tissue levels promote endothelial dysfunction

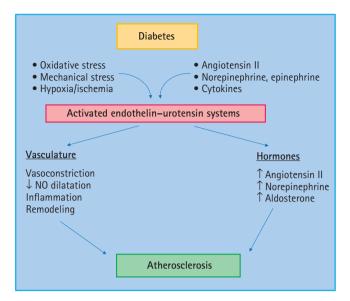


Figure 39.8 Proposed mechanisms of the endothelin and urotensin systems on atherosclerosis in diabetes. In diabetes, activation of the endothelin system induces vasoconstriction, vascular smooth muscle cells proliferation, wall thickening, inflammation and tissue remodeling thus leading to the development and progression of atherosclerosis.

and vascular structural alterations *via* the activation of ET_A receptors [163,165]. Treatment with an ET_A receptor antagonist decreased atherosclerotic lesion area in the aorta in apoE KO mice [11]. Similar results have been reported with a non-selective ET_A/ET_B receptor blocker in rabbits. The role of the ET_B receptor in atherosclerosis remains controversial at this stage although it has been shown to have anti-atherosclerotic effects via stimulation of NO production [166].

Finally, some of the beneficial effects of ACE-inhibitors in term of cardiovascular protection may be at least in part mediated by ET inhibition. Captopril inhibits the release of ET from cultured human endothelial cells and decreases 24-h urinary ET-1 excretion [167,168]. Furthermore, ET_A receptor antagonism appears to be beneficial in the treatment of diabetic nephropathy, but there are currently no data to support a protective effect on macrovascular disease in people with diabetes.

Urotensin II

Several recent reports have revealed the powerful vasocontractile effect of urotensin II (UT II) consistent with a potential importance of this peptide in cardiovascular physiology and diseases (Figure 39.8) [169]. A specific UT II receptor was identified in 1999 [170]. Elevated circulating concentrations of UT II have been detected in patients with various CVD states including heart failure, hypertension [171], carotid atherosclerosis, pre-eclampsia and eclampsia, renal dysfunction and diabetes mellitus [172]. In addition, upregulation of the UT receptor system has been found in patients with congestive heart failure and pulmonary hypertension [173].

Role of urotensin II in atherosclerosis

Expression of UT II is upregulated in endothelial, myointimal and medial smooth muscle cells of atherosclerotic human coronary arteries. Bousette *et al.* [12] have demonstrated that UT II expression is increased in both atherosclerotic carotid arteries and aortas. It has been shown that the plasma UT II level is correlated positively with carotid atherosclerosis in patients with essential hypertension [174]. In apoE KO mice, UT expression was significantly higher than in wild-type control mice [175]. Chronic infusion of UT II enhances atherosclerotic lesion in the aorta of apoE KO mice by increasing ROS production and acyl-CoA-cholesterol acyl-transferase 1 expression. UT II expression in endothelial cells and VSMCs is increased following balloon injury in rat carotid arteries [176]. Furthermore, treatment with a UT II receptor blocker was associated with a 60% reduction in intima lesion development.

UT II is linked to the activation of the redox-sensitive enzyme NADPH oxidase in the vascular wall and increased expression of inflammatory cytokines (Figure 39.8). In human peripheral blood mononuclear cells, inflammatory stimuli including IL-1 β and TNF- α strongly enhance UT receptor mRNA and protein expression [177]. Of direct relevance to atherosclerosis, UT II has been shown, predominantly in *in vitro* studies to enhance LDL cholesterot and ROS production via NADPH oxidase and to promote monocyte recruitment.

TNF-related apoptosis inducing ligand and osteoprotegerin

Tumor necrosis factor (TNF) related apoptosis inducing ligand (TRAIL) is a member of the TNF ligand family [178]. Both the membrane-bound and soluble forms of TRAIL rapidly induce apoptosis in multiple transformed cell lines and tumor cells, but not in normal cells [178,179].

Osteoprotegerin (OPG) is a member of TNF-receptor superfamily [180], which is produced by a variety of tissues including the cardiovascular system. OPG has two known TNF family ligands: receptor activator of NF κ B ligand (RANKL) [181] and TRAIL [182]. Increasing experimental evidence suggests that both TRAIL and OPG are involved in vascular pathophysiology.

Recently, the potential role of soluble recombinant (sr) TRAIL in the pathogenesis and/or treatment of diabetes-induced atherosclerosis has been investigated *in vivo*, in streptozotocin-diabetic apoE KO mice [183]. Repeated intraperitoneal injections of srTRAIL significantly attenuated plaque development and contributed to stabilize atherosclerotic lesions by selectively decreasing the number of infiltrating macrophages and increasing the VSMCs within the atherosclerotic plaques [183]. The potential clinical relevance of these results is corroborated by two studies in patients with acute coronary syndrome, revealing significantly lower sTRAIL serum levels in these subjects when compared to patients with stable angina or normal coronary arteries [184,185].

OPG-deficient mice demonstrated calcifications of the aorta and renal arteries [186], suggesting that OPG might have a role in protecting from vascular calcification. Interestingly, a study in elderly women found a significant correlation between elevated OPG serum levels and cardiovascular mortality [187]. The potential links between OPG and vascular disease in humans have been further supported by the detection of a single nucleotide polymorphism in the promoter region of the human gene for OPG related to vascular morphology and function [188]. Several independent studies have reported elevated serum and/or plasma OPG levels in people with diabetes, and in particular in people with diabetes and vascular complications [189]. Although some authors have proposed that the increase in OPG levels may represent a defence mechanism against other factors that promote vascular pathologies, other studies support a pro-atherosclerotic role for OPG in the vasculature [190].

Complement activation

The complement system consists of approximately 30 proteins that provide an important component of the innate immune system. The complement system is activated through three different pathways: the classic, the alternative and the mannosebinding lectin (MBL) complement pathway, respectively. These three pathways merge and stimulate the formation of a C3 convertase (Figure 39.9).

Recently, an increasing amount of evidence indicates that the complement system activation may have a role in the development of diabetic macrovascular complications [15], in particular

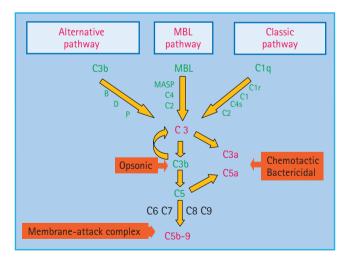


Figure 39.9 Schematic presentation of the complement system. The complement cascade is activated by either of the three pathways: the classic, the mannose-binding lectin (MBL) pathway, or the alternative pathway. Activation leads to formation of the membrane-attack complex as well as formation of opsonic, chemotactic and bactericidal factors.

the MBL pathway as the initiating pathway of complement activation in patients with diabetes [15]. Patients with T1DM with nephropathy and CVD have significantly higher circulating levels of MBL than normoalbuminuric patients [191]. Furthermore, there is evidence that MBL levels may have prognostic relevance in people with diabetes. Several observations have suggested that MBL may be involved in the pathogenesis of microvascular and macrovascular complications in T1DM and that determination of MBL status might be useful to identify patients at increased risk of developing these complications [191,192]. High levels of MBL early in the course of T1DM are significantly associated with later development of persistent microalbuminuria and macroalbuminuria [193]. In addition, in patients with T2DM, measurement of MBL has been shown to provide prognostic information on mortality and development of microalbuminuria [194]. The mechanism responsible for this potential activation of the complement system in patients with diabetes is still unknown. It has been hypothesized that hyperglycemia in people with diabetes may activate the complement system via mitochondrial ROS production or enhanced RAGE activation.

Clinical studies using pexelizumab, an inhibitor of C5, have recently demonstrated a significant reduction in mortality following acute MI as well as reducing the incidence of death or MI in patients after coronary artery bypass graft surgery [195]. Further studies are needed to determine in particular if manipulation of the complement system may represent a promising target in the treatment and prevention of diabetes associated macrovascular disease.

Interventions to reduce diabetes associated macrovascular complications

A range of interventions has been proposed to reduce the macrovascular complications of diabetes including lifestyle modification, weight reduction, dietary changes and regular exercise.

Glucose control

Optimal glucose control is pivotal for the prevention and treatment of microvascular complications of diabetes; however, based on the recent clinical trials [22,23], the evidence for beneficial effects on macrovascular outcomes is less clear. Nevertheless, it has been suggested to aim for a HbA_{1c} <7% (53 mmol/mol), but to be more cautious in patients with underlying and pre-existing CVD.

Hypertension

Epidemiologic studies have shown that the risk for cardiovascular events and mortality starts at BP values as low as 115/75 mmHg for the general population and doubles for every increase in 20 mmHg systolic and 10 mmHg diastolic (see Chapter 40). The question of what systolic or diastolic BP level should be targeted has not been completely answered by currently available outcome trials. Based on studies such as the Hypertension Optimal Treatment (HOT) study [196], UKPDS [197] and the Appropriate Blood Pressure Control in type 2 Diabetes (ABCD) trial [198], a maximum systolic BP of 130–135 mmHg should be the goal for people with diabetes.

The more recently completed ADVANCE trial [23] evaluated the effect of a fixed dose of perindopril and indapamide on macrovascular and microvascular outcomes in subjects with T2DM. The treatment group showed a mean reduction in systolic and diastolic BP of 5.6 and 2.2 mmHg respectively, which was associated with a 18% reduction in cardiovascular deaths and a 14% reduction in coronary events in comparison to the control group. No lower limit for BP reduction appears to exist at which benefits related to cardiovascular outcomes are not observed with a similar pattern recently reported for renal disease [199]. The ACCORD trial is currently testing the effects of lowering BP below 140/90 mmHg on cardiovascular outcomes. Specifically, it will evaluate if a BP reduction to 120 mmHg systolic will further reduce the cardiovascular event rate.

Choice of antihypertensive treatment

Experimental studies have clearly demonstrated a reduction in atherosclerosis with RAS inhibitors with a possible superiority of ACE inhibitors compared to ARBs in T2DM; however, in the human setting, there are no conclusive studies as yet to show clear superiority of one class of antihypertensive drugs over another. The majority of the beneficial effects on cardiovascular events appear to be related to the antihypertensive actions of these agents. The ONTARGET study was the first study to evaluate the effect of combined RAS blockade in comparison to full-dose ramipril and telmisartan alone [200]. There was no superiority of the ACE inhibitor versus the ARB in terms of cardiovascular events, and dual blockade did not confer greater cardiovascular protection and indeed was associated with more adverse events; however, only 30% of all subjects enrolled in this study had diabetes, and no specific subgroup analysis has been reported so far. Therefore, this study cannot conclusively answer the question of superiority of different blockers of the RAS and combination RAS-blockade on microvascular and macrovascular outcomes in diabetes.

Dyslipidemia

The lipid profile is usually altered in the diabetic milieu and further altered in the context of liver or renal disease. Dyslipidemia further accelerates atherosclerosis development and progression (see Chapter 40). At any given level of cholesterol, a person with diabetes has a two- to threefold increased cardiovascular risk than a person without diabetes.

Current guidelines based on numerous studies particularly with statins (Heart Protective Study [HPS] [201], the Collaborative Atorvastatin Diabetes Study [CARDS] [202]) suggest to reduce LDL levels below 2.5 mmol/L with some more recent evidence suggesting enhanced benefits if LDL is lowered <2 mmol/L, triglycerides <1 mmol/L and HDL> 1 mmol/L according to preexisting CVD and risk [203].

Hypercoagulability

Diabetes is associated with pro-thrombotic changes and enhanced coagulability, thus increasing cardiovascular risk [204,205].

Although not studied in detail, agents such as aspirin may be useful in people with diabetes, with increasing evidence that higher doses of these agents are often required in people with diabetes to confer cardiovascular benefits. Other drugs to consider in diabetes are clopidogrel as well as the use of drugs such as abciximab [206], which has been reported to be particularly effective in people with diabetes.

Novel therapies

Several approaches have been used to intervene in diabetes-associated complications including aldose reductase inhibitors, PKC isoform inhibitors, in particular PKC β , but most of the evidence for a potential role for these agents has been obtained in the context of microvascular complications. Furthermore, inhibitors of glucose induced ROS formation, such as benfotiamine, have been investigated in clinical trials with variable results. AGE inhibitors and RAGE antagonism have also been employed, mainly in the context of diabetic nephropathy. The cross-link breaker, alagebrium, has shown to reduce arterial stiffness in hypertensive patients and to improve left ventricular function. Direct antagonists of RAGE and human chimeric sRAGE are also currently being investigated in early clinical trials, however, not as yet with respect to macrovascular disease.

The role of PPAR- α and PPAR- γ agonists have also been investigated with modest or no effects on cardiovascular outcomes [207–209]. The dual PPAR- α/γ agonists such as muraglitazar do not appear to be likely to be introduced as a result of adverse side effects and the possibility of adverse cardiovascular events [210].

Multifactorial approaches

The Steno-2 study, using a multifactorial approach addressing lipids, BP and hyperglycemia, demonstrated significant improvements on cardiovascular outcomes [25]. Thus, a synergistic multifactorial approach addressing glycemic control in the context of additional strategies to reduce concomitant cardiovascular risk factors remains the best approach to be currently considered. Furthermore, strategies directed towards a reduction in activation of vasoactive systems such as the RAS, the ET and UT systems, the AGE/RAGE axis, novel proteins such as TRAIL and the complement system as well as oxidative stress and inflammation may additional promising approaches to prevent and minimize macrovascular disease in diabetes.

References

- 1 Nathan DM, Lachin J, Cleary P, Orchard T, Brillon DJ, Backlund JY, *et al.* Intensive diabetes therapy and carotid intima-media thickness in type 1 diabetes mellitus. *N Engl J Med* 2003; **348**: 2294–2303.
- 2 Nathan DM, Cleary PA, Backlund JY, Genuth SM, Lachin JM, Orchard TJ, *et al.* Intensive diabetes treatment and cardiovascular disease in patients with type 1 diabetes. *N Engl J Med* 2005; 353:2643–2653.

- 3 Turner RC, Millns H, Neil HA, Stratton IM, Manley SE, Matthews DR, *et al.* Risk factors for coronary artery disease in non-insulin dependent diabetes mellitus: United Kingdom Prospective Diabetes Study (UKPDS 23). *Br Med J* 1998; **316**:823–828.
- 4 Turner R, Cull C, Holman R. UK Prospective Diabetes Study 17: a 9-year update of a randomized, controlled trial on the effect of improved metabolic control on complications in non-insulindependent diabetes mellitus. *Ann Intern Med* 1996; **124**:136–145.
- 5 Holman RR, Paul SK, Bethel MA, Neil HA, Matthews DR. Longterm follow-up after tight control of blood pressure in type 2 diabetes. *N Engl J Med* 2008; **359**:1565–1576.
- 6 Califf RM, Boolell M, Haffner SM, Bethel MA, McMurray J, Duggal A, *et al.* Prevention of diabetes and cardiovascular disease in patients with impaired glucose tolerance: rationale and design of the Nateglinide and Valsartan in Impaired Glucose Tolerance Outcomes Research (NAVIGATOR) Trial. *Am Heart J* 2008; **156**:623–632.
- 7 Stamler J, Vaccaro O, Neaton JD, Wentworth D. Diabetes, other risk factors, and 12-yr cardiovascular mortality for men screened in the Multiple Risk Factor Intervention Trial. *Diabetes Care* 1993; **16**:434–444.
- 8 Fitzgerald AP, Jarrett RJ. Are conventional risk factors for mortality relevant in type 2 diabetes? *Diabet Med* 1991; **8**:475–480.
- 9 Dale AC, Vatten LJ, Nilsen TI, Midthjell K, Wiseth R. Secular decline in mortality from coronary heart disease in adults with diabetes mellitus: cohort study. *Br Med J* 2008; **337**:a236.
- 10 Calcutt NA, Cooper ME, Kern TS, Schmidt AM. Therapies for hyperglycaemia-induced diabetic complications: from animal models to clinical trials. *Nat Rev Drug Discov* 2009; 8:417–429.
- 11 Barton M, Haudenschild CC, d'Uscio LV, Shaw S, Munter K, Luscher TF. Endothelin ETA receptor blockade restores NOmediated endothelial function and inhibits atherosclerosis in apolipoprotein E-deficient mice. *Proc Natl Acad Sci U S A* 1998; **95**:14367–14372.
- 12 Bousette N, Patel L, Douglas SA, Ohlstein EH, Giaid A. Increased expression of urotensin II and its cognate receptor GPR14 in atherosclerotic lesions of the human aorta. *Atherosclerosis* 2004; 176:117–123.
- 13 Hassan GS, Douglas SA, Ohlstein EH, Giaid A. Expression of urotensin-II in human coronary atherosclerosis. *Peptides* 2005; 26:2464–2472.
- 14 Secchiero P, Zerbinati C, Rimondi E, Corallini F, Milani D, Grill V, *et al.* TRAIL promotes the survival, migration and proliferation of vascular smooth muscle cells. *Cell Mol Life Sci* 2004; **61**: 1965–1974.
- 15 Østergaard J, Hansen TK, Thiel S, Flyvbjerg A. Complement activation and diabetic vascular complications. *Clin Chim Acta* 2005; 361:10–19.
- 16 Libby P. Current concepts of the pathogenesis of the acute coronary syndromes. *Circulation* 2001; 104:365–372.
- 17 UK Prospective Diabetes Study (UKPDS) Group. Intensive bloodglucose control with sulphonylureas or insulin compared with conventional treatment and risk of complications in patients with type 2 diabetes (UKPDS 33). *Lancet* 1998; **352**:837–853.
- 18 Gall MA, Borch-Johnsen K, Hougaard P, Nielsen FS, Parving H-H. Albuminuria and poor glycemic control predict mortality in NIDDM. *Diabetes* 1995; 44:1303–1309.
- 19 Salomaa V, Miettinen H, Kuulasmaa K, Niemelä M, Ketonen M, Vuorenmaa T, *et al.* Decline of coronary heart disease mortality in Finland during 1983 to 1992: roles of incidence, recurrence, and

case-fatality. The FINMONICA MI Register Study. *Circulation* 1996; **94**:3130–3137.

- 20 Kuusisto J, Mykkanen L, Pyorala K, Laakso M. Non-insulindependent diabetes and its metabolic control are important predictors of stroke in elderly subjects. *Stroke* 1994; 25:1157–1164.
- 21 Gerstein HC. Is glucose a continuous risk factor for cardiovascular mortality? *Diabetes Care* 1999; **22**:659–660.
- 22 Gerstein HC, Miller ME, Byington RP, Goff DC Jr, Bigger JT, Buse JB, *et al.* Effects of intensive glucose lowering in type 2 diabetes. *N Engl J Med* 2008; **358**:2545–2559.
- 23 Patel A, MacMahon S, Chalmers J, Neal B, Billot L, Woodward M, Marre M, *et al.*; ADVANCE Collaborative Group. Intensive blood glucose control and vascular outcomes in patients with type 2 diabetes. *N Engl J Med* 2008; **358**:2560–2572.
- 24 Duckworth W, Abraira C, Moritz T, Reda D, Emanuele N, Reaven PD, *et al.* Glucose control and vascular complications in veterans with type 2 diabetes. *N Engl J Med* 2009; **360**:129–139.
- 25 Gæde P, Lund-Andersen H, Parving H-H, Pedersen O. Effect of a multifactorial intervention on mortality in type 2 diabetes. N Engl J Med 2008; 358:580–591.
- 26 Holman RR, Paul SK, Bethel MA, Matthews DR, Neil HA. 10-year follow-up of intensive glucose control in type 2 diabetes. *N Engl J Med* 2008; **359**:1577–1589.
- 27 Vikramadithyan RK, Hu Y, Noh HL, Liang CP, Hallam K, Tall AR, *et al.* Human aldose reductase expression accelerates diabetic atherosclerosis in transgenic mice. *J Clin Invest* 2005; **115**: 2434–2443.
- 28 Brownlee M. Biochemistry and molecular cell biology of diabetic complications. *Nature* 2001; 414:813–820.
- 29 Hsueh W, Abel ED, Breslow JL, Maeda N, Davis RC, Fisher EA, et al. Recipes for creating animal models of diabetic cardiovascular disease. *Circ Res* 2007; 100:1415–1427.
- 30 Renard CB, Kramer F, Johansson F, Lamharzi N, Tannock LR, von Herrath MG, et al. Diabetes and diabetes-associated lipid abnormalities have distinct effects on initiation and progression of atherosclerotic lesions. J Clin Invest 2004; 114:659–668.
- 31 Johansson F, Kramer F, Barnhart S, Kanter JE, Vaisar T, Merrill RD, et al. Type 1 diabetes promotes disruption of advanced atherosclerotic lesions in LDL receptor-deficient mice. Proc Natl Acad Sci U S A 2008; 105:2082–2087.
- 32 Brownlee M, Hirsch IB. Glycemic variability: a hemoglobin A1cindependent risk factor for diabetic complications. *JAMA* 2006; 295:1707–1708.
- 33 Monnier L, Mas E, Ginet C, Michel F, Villon L, Cristol JP, *et al.* Activation of oxidative stress by acute glucose fluctuations compared with sustained chronic hyperglycemia in patients with type 2 diabetes. *JAMA* 2006; **295**:1681–1687.
- 34 El-Osta A, Brasacchio D, Yao D, Pocai A, Jones PL, Roeder RG, *et al.* Transient high glucose causes persistent epigenetic changes and altered gene expression during subsequent normoglycemia. *J Exp Med* 2008; **205**:2409–2417.
- 35 Brasacchio D, Okabe J, Tikellis C, Balcerczyk A, George P, Baker EK, et al. Hyperglycemia induces a dynamic cooperativity of histone methylase and demethylase enzymes associated with gene-activating epigenetic marks that coexist on the lysine tail. *Diabetes* 2009; 58:1229–1236.
- 36 Saydah SH, Loria CM, Eberhardt MS, Brancati FL. Subclinical states of glucose intolerance and risk of death in the US. *Diabetes Care* 2001; 24:447–453.

- 37 Haffner S. Mortality from coronary heart disease in subjects with type 2 diabetes and in nondiabetic subjects with and without prior myocardial infarction. *N Engl J Med* 1998; **339**:229–234.
- 38 Hu G, Qiao Q, Tuomilehto J, Balkau B, Borch-Johnsen K, Pyorala K. Prevalence of the metabolic syndrome and its relation to all-cause and cardiovascular mortality in nondiabetic European men and women. Arch Intern Med 2004; 164:1066–1076.
- 39 Yip J, Facchini FS, Reaven GM. Resistance to insulin-mediated glucose disposal as a predictor of cardiovascular disease. J Clin Endocrinol Metab 1998; 83:2773–2776.
- 40 Hanley AJ, Williams K, Stern MP, Haffner SM. Homeostasis model assessment of insulin resistance in relation to the incidence of cardiovascular disease: the San Antonio Heart Study. *Diabetes Care* 2002; 25:1177–1184.
- 41 Hanley AJ, McKeown-Eyssen G, Harris SB, Hegele RA, Wolever TM, Kwan J, *et al.* Association of parity with risk of type 2 diabetes and related metabolic disorders. *Diabetes Care* 2002; **25**:690–695.
- 42 Hsueh WA, Law RE. Cardiovascular risk continuum: implications of insulin resistance and diabetes. *Am J Med* 1998; **105**:4S–14S.
- 43 King GL, Brownlee M. The cellular and molecular mechanisms of diabetic complications. *Endocrinol Metab Clin North Am* 1996; 25:255–270.
- 44 Yuen DY, Dwyer RM, Matthews VB, Zhang L, Drew BG, Neill B, *et al.* Interleukin-6 attenuates insulin-mediated increases in endothelial cell signaling but augments skeletal muscle insulin action via differential effects on tumor necrosis factor-alpha expression. *Diabetes* 2009; **58**:1086–1095.
- 45 Lo TW, Westwood ME, McLellan AC, Selwood T, Thornalley PJ. Binding and modification of proteins by methylglyoxal under physiological conditions: a kinetic and mechanistic study with N alphaacetylarginine, N alpha-acetylcysteine, and N alpha-acetyllysine, and bovine serum albumin. J Biol Chem 1994; 269:32299–32305.
- 46 Ahmed N, Thornalley PJ, Dawczynski J, Franke S, Strobel J, Stein G, et al. Methylglyoxal-derived hydroimidazolone advanced glycation end-products of human lens proteins. *Invest Ophthalmol Vis Sci* 2003; 44:5287–5292.
- 47 Thornalley PJ, Langborg A, Minhas HS. Formation of glyoxal, methylglyoxal and 3-deoxyglucosone in the glycation of proteins by glucose. *Biochem J* 1999; 344:109–116.
- 48 Brownlee M. Advanced protein glycosylation in diabetes and aging. Annu Rev Med 1995; 46:223–234.
- 49 Vlassara H, Fuh H, Makita Z, Krungkrai S, Cerami A, Bucala R. Exogenous advanced glycosylation end products induce complex vascular dysfunction in normal animals: a model for diabetic and aging complications. *Proc Natl Acad Sci U S A* 1992; 89:12043–12047.
- 50 Bucala R, Tracey KJ, Cerami A. Advanced glycosylation products quench nitric oxide and mediate defective endothelium-dependent vasodilatation in experimental diabetes. *J Clin Invest* 1991; 87:432–438.
- 51 Forbes JM, Yee LT, Thallas V, Lassila M, Candido R, Jandeleir-Dahm KA, *et al.* Advanced glycation end product interventions reduce diabetes-accelerated atherosclerosis. *Diabetes* 2004; 53:1813–1823.
- 52 Kislinger T, Tanji N, Wendt T, Qu W, Lu Y, Ferran LJ Jr, et al. Receptor for advanced glycation end products mediates inflammation and enhanced expression of tissue factor in vasculature of diabetic apolipoprotein E-null mice. Arterioscler Thromb Vasc Biol 2001; 21:905–910.

- 53 Aronson D. Cross-linking of glycated collagen in the pathogenesis of arterial and myocardial stiffening of aging and diabetes. *J Hypertens* 2003; **21**:3–12.
- 54 Kass DA, Shapiro EP, Kawaguchi M, Capriotti AR, Scuteri A, deGroof RC, *et al.* Improved arterial compliance by a novel advanced glycation end-product crosslink breaker. *Circulation* 2001; 104:1464–470.
- 55 Hogan M, Cerami A, Bucala R. Advanced glycosylation endproducts block the antiproliferative effect of nitric oxide: role in the vascular and renal complications of diabetes mellitus. *J Clin Invest* 1992; 90:1110–1115.
- 56 Wautier MP, Chappey O, Corda S, Stern DM, Schmidt AM, Wautier JL. Activation of NADPH oxidase by AGE links oxidant stress to altered gene expression via RAGE. Am J Physiol Endocrinol Metab 2001; 280:E685–694.
- 57 Neeper M, Schmidt AM, Brett J, Yan SD, Wang F, Pan YC, *et al.* Cloning and expression of a cell surface receptor for advanced glycosylation end products of proteins. *J Biol Chem* 1992; **267**:14998–15004.
- 58 McRobert EA, Gallicchio M, Jerums G, Cooper ME, Bach LA. The amino-terminal domains of the ezrin, radixin, and moesin (ERM) proteins bind advanced glycation end products, an interaction that may play a role in the development of diabetic complications. *J Biol Chem* 2003; 278:25783–25789.
- 59 Ohgami N, Nagai R, Ikemoto M, Arai H, Kuniyasu A, Horiuchi S, et al. CD36, a member of the class b scavenger receptor family, as a receptor for advanced glycation end products. J Biol Chem 2001; 276:3195–3202.
- 60 Febbraio M, Guy E, Silverstein RL. Stem cell transplantation reveals that absence of macrophage CD36 is protective against atherosclerosis. *Arterioscler Thromb Vasc Biol* 2004; **24**:2333–2338.
- 61 Soulis T, Thallas V, Youssef S, Gilbert RE, McWilliam BG, Murray-McIntosh RP, *et al.* Advanced glycation end products and their receptors co-localize in rat organs susceptible to diabetic microvascular injury. *Diabetologia* 1997; **40**:619–628.
- 62 Brett J, Schmidt AM, Yan SD, Zou YS, Weidman E, Pinsky D, *et al.* Survey of the distribution of a newly characterized receptor for advanced glycation end products in tissues. *Am J Pathol* 1993; **143**:1699–1712.
- 63 Bierhaus A, Stern DM, Nawroth PP. RAGE in inflammation: a new therapeutic target? *Curr Opin Investig Drugs* 2006; 7:985–991.
- 64 Schmidt AM, Yan SD, Yan SF, Stern DM. The multiligand receptor RAGE as a progression factor amplifying immune and inflammatory responses. *J Clin Invest* 2001; **108**:949–955.
- 65 Lander HM, Tauras JM, Ogiste JS, Hori O, Moss RA, Schmidt AM. Activation of the receptor for advanced glycation end products triggers a p21(ras)-dependent mitogen-activated protein kinase pathway regulated by oxidant stress. *J Biol Chem* 1997; **272**:17810–17814.
- 66 Li JH, Wang W, Huang XR, Oldfield M, Schmidt AM, Cooper ME, et al. Advanced glycation end products induce tubular epithelialmyofibroblast transition through the RAGE-ERK1/2 MAP kinase signaling pathway. Am J Pathol 2004; 164:1389–1397.
- 67 Li J, Schmidt AM. Characterization and functional analysis of the promoter of RAGE, the receptor for advanced glycation end products. J Biol Chem 1997; 272:16498–16506.
- 68 Higashi T, Sano H, Saishoji T, Ikeda K, Jinnouchi Y, Kanzaki T, et al. The receptor for advanced glycation end products mediates the chemotaxis of rabbit smooth muscle cells. *Diabetes* 1997; 46:463–472.

- 69 Miyata T, Ueda Y, Asahi K, Izuhara Y, Inagi R, Saito A, *et al.* Mechanism of the inhibitory effect of OPB-9195 ((+/–)-2isopropylidenehydrazono-4-oxo-thiazolidin-5-yla cetanilide) on advanced glycation end product and advanced lipoxidation end product formation. *J Am Soc Nephrol* 2000; **11**:1719–1725.
- 70 Thornalley PJ, Yurek-George A, Argirov OK. Kinetics and mechanism of the reaction of aminoguanidine with the alphaoxoaldehydes glyoxal, methylglyoxal, and 3-deoxyglucosone under physiological conditions. *Biochem Pharmacol* 2000; 60:55–65.
- 71 Rosca MG, Mustata TG, Kinter MT, Ozdemir AM, Kern TS, Szweda LI, *et al.* Glycation of mitochondrial proteins from diabetic rat kidney is associated with excess superoxide formation. *Am J Physiol Renal Physiol* 2005; 289:F420–430.
- 72 Soulis-Liparota T, Cooper M, Papazoglou D, Clarke B, Jerums G. Retardation by aminoguanidine of development of albuminuria, mesangial expansion, and tissue fluorescence in streptozocininduced diabetic rat. *Diabetes* 1991; **40**:1328–1334.
- 73 Coughlan MT, Thallas-Bonke V, Pete J, Long DM, Gasser A, Tong DC, *et al.* Combination therapy with the advanced glycation end product cross-link breaker, alagebrium, and angiotensin converting enzyme inhibitors in diabetes: synergy or redundancy? *Endocrinology* 2007; 148:886–895.
- 74 Vasan S, Zhang X, Kapurniotu A, Bernhagen J, Teichberg S, Basgen J, et al. An agent cleaving glucose-derived protein crosslinks in vitro and in vivo. Nature 1996; 382:275–278.
- 75 Asif M, Egan J, Vasan S, Jyothirmayi GN, Masurekar MR, Lopez S, et al. An advanced glycation endproduct cross-link breaker can reverse age-related increases in myocardial stiffness. Proc Natl Acad Sci U S A 2000; 97:2809–2813.
- 76 Wolffenbuttel BH, Boulanger CM, Crijns FR, Huijberts MS, Poitevin P, Swennen GN, *et al.* Breakers of advanced glycation end products restore large artery properties in experimental diabetes. *Proc Natl Acad Sci U S A* 1998; **95**:4630–4634.
- 77 Candido R, Forbes JM, Thomas MC, Thallas V, Dean RG, Burns WC, *et al.* A breaker of advanced glycation end products attenuates diabetes-induced myocardial structural changes. *Circ Res* 2003; 92:785–792.
- 78 Lassila M, Seah KK, Allen TJ, Thallas V, Thomas MC, Candido R, et al. Accelerated nephropathy in diabetic apolipoprotein e-knockout mouse: role of advanced glycation end products. J Am Soc Nephrol 2004; 15:2125–2138.
- 79 Forbes JM, Thallas V, Thomas MC, Founds HW, Burns WC, Jerums G, et al. The breakdown of preexisting advanced glycation end products is associated with reduced renal fibrosis in experimental diabetes. *Faseb J* 2003; 17:1762–1764.
- 80 Thallas-Bonke V, Lindschau C, Rizkalla B, Bach LA, Boner G, Meier M, et al. Attenuation of extracellular matrix accumulation in diabetic nephropathy by the advanced glycation end product cross-link breaker ALT-711 via a protein kinase C-alpha-dependent pathway. Diabetes 2004; 53:2921–2930.
- 81 Burns WC, Twigg SM, Forbes JM, Pete J, Tikellis C, Thallas-Bonke V, et al. Connective tissue growth factor plays an important role in advanced glycation end product-induced tubular epithelial-to-mesenchymal transition: implications for diabetic renal disease. J Am Soc Nephrol 2006; 17:2484–2494.
- 82 Twigg SM, Cao Z, MCLennan SV, Burns WC, Brammar G, Forbes JM, *et al.* Renal connective tissue growth factor induction in experimental diabetes is prevented by aminoguanidine. *Endocrinology* 2002; 143:4907–4915.

- 83 Yonekura H, Yamamoto Y, Sakurai S, Petrova RG, Abedin MJ, Li H, *et al.* Novel splice variants of the receptor for advanced glycation end-products expressed in human vascular endothelial cells and pericytes, and their putative roles in diabetes-induced vascular injury. *Biochem J* 2003; **370**:1097–1109.
- 84 Hanford LE, Enghild JJ, Valnickova Z, Petersen SV, Schaefer LM, Schaefer TM, *et al.* Purification and characterization of mouse soluble receptor for advanced glycation end products (sRAGE). *J Biol Chem* 2004; 279:50019–50024.
- 85 Park L, Raman KG, Lee KJ, Ferran LJ Jr, Chow WS, Stern D, et al. Suppression of accelerated diabetic atherosclerosis by the soluble receptor for advanced glycation endproducts. *Nat Med* 1998; 4:1025–1031.
- 86 Bucciarelli LG, Wendt T, Qu W, Lu Y, Lalla E, Rong LL, et al. RAGE blockade stabilizes established atherosclerosis in diabetic apolipoprotein E-null mice. *Circulation* 2002; 106:2827–2835.
- 87 Naka Y, Bucciarelli LG, Wendt T, Lee LK, Rong LL, Ramasamy R, et al. RAGE axis: animal models and novel insights into the vascular complications of diabetes. *Arterioscler Thromb Vasc Biol* 2004; 24:1342–1349.
- 88 Wendt T, Harja E, Bucciarelli L, Qu W, Lu Y, Rong LL, et al. RAGE modulates vascular inflammation and atherosclerosis in a murine model of type 2 diabetes. *Atherosclerosis* 2006; 185:70–77.
- 89 Wendt TM, Tanji N, Guo J, Kislinger TR, Qu W, Lu Y, et al. RAGE drives the development of glomerulosclerosis and implicates podocyte activation in the pathogenesis of diabetic nephropathy. Am J Pathol 2003; 162:1123–1137.
- 90 Soro-Paavonen A, Watson AM, Li J, paavonen K, Koitka A, Calkin AC, et al. Receptor for advanced glycation end products (RAGE) deficiency attenuates the development of atherosclerosis in diabetes. *Diabetes* 2008; 57:2461–2469.
- Cooper ME. Interaction of metabolic and hemodynamic factors in mediating experimental diabetic nephropathy. *Diabetologia* 2001; 44:1957–1972.
- 92 Thomas MC, Tikellis C, Burns WM, Bialkowski K, Cao Z, Coughlan MT, et al. Interactions between renin angiotensin system and advanced glycation in the kidney. J Am Soc Nephrol 2005; 16:2976–2984.
- 93 Forbes JM, Thorpe SR, Thallas-Bonke V, Pete J, Thomas MC, Deemer ER, et al. Modulation of soluble receptor for advanced glycation end products by angiotensin-converting enzyme-1 inhibition in diabetic nephropathy. J Am Soc Nephrol 2005; 16:2363–2372.
- 94 Calkin AC, Cooper ME, Jandeleit-Dahm KA, Allen TJ. Gemfibrozil decreases atherosclerosis in experimental diabetes in association with a reduction in oxidative stress and inflammation. *Diabetologia* 2006; 49:766–774.
- 95 Yoshida T, Yamagishi S, Nakamura K, Matsui T, Imaizumi T, Takeuchi M, *et al.* Telmisartan inhibits AGE-induced C-reactive protein production through downregulation of the receptor for AGE via peroxisome proliferator-activated receptor-gamma activation. *Diabetologia* 2006; **49**:3094–3099.
- 96 Cuccurullo C, Iezzi A, Fazia ML, De Cesare D, Di Francesco A, Muraro R, et al. Suppression of RAGE as a basis of simvastatindependent plaque stabilization in type 2 diabetes. Arterioscler Thromb Vasc Biol 2006; 26:2716–2723.
- 97 Tigerstedt R, Bergman PG. Niere und Kreislauf. *Scand Arch Physiol* 1898; **8**:223–228.

- 98 Admiraal PJ, Derkx FH, Danser AH, Pieterman H, Schalekamp MA. Metabolism and production of angiotensin I in different vascular beds in subjects with hypertension. *Hypertension* 1990; 15: 44–55.
- 99 Donoghue M, Hsieh F, Baronas E, Godbout K, Gosselin M, Stagliano N, et al. A novel angiotensin-converting enzyme-related carboxy-peptidase (ACE2) converts angiotensin I to angiotensin 1-9. Circ Res 2000; 87:E1–9.
- 100 Ferrario CM. Angiotensin-converting enzyme 2 and angiotensin-(1-7): an evolving story in cardiovascular regulation. *Hypertension* 2006; 47:515–521.
- 101 Stoll M, Steckelings UM, Paul M, Bottari SP, Metzger R, Unger T. The angiotensin AT2-receptor mediates inhibition of cell proliferation in coronary endothelial cells. J Clin Invest 1995; 95:651–657.
- 102 Candido R, Jandeleit-Dahm KA, Cao Z, Nesteroff SP, Burns WC, Twigg SM, *et al.* Prevention of accelerated atherosclerosis by angiotensin-converting enzyme inhibition in diabetic apolipoprotein E-deficient mice. *Circulation* 2002; **106**:246–253.
- 103 Schmieder RE, Hilgers KF, Schlaich MP, Schmidt BM. Reninangiotensin system and cardiovascular risk. *Lancet* 2007; 369:1208–1219.
- 104 Diet F, Pratt RE, Berry GJ, Momose N, Gibbons GH, Dzau VJ. Increased accumulation of tissue ACE in human atherosclerotic coronary artery disease. *Circulation* 1996; **94**:2756–2767.
- 105 Hansson L, Lindholm LH, Niskanen L, Lanke J, Hedner T, Niklason A, Luomanmäki K, et al. Effect of angiotensin-converting-enzyme inhibition compared with conventional therapy on cardiovascular morbidity and mortality in hypertension: the Captopril Prevention Project (CAPP) randomised trial. *Lancet* 1999; 353:611–616.
- 106 Yusuf S, Ostergren JB, Gerstein HC, Pfeffer MA, Swedberg K, Granger CB, et al.; Candesartan in Heart Failure-Assessment of Reduction in Mortality and Morbidity Program Investigators. Effects of candesartan on the development of a new diagnosis of diabetes mellitus in patients with heart failure. *Circulation* 2005; 112:48–53.
- 107 Lindholm LH, Ibsen H, Borch-Johnsen K, Olsen MH, Wachtell K, Dahlöf B, *et al.* Risk of new-onset diabetes in the Losartan Intervention For Endpoint reduction in hypertension study. *J Hypertens* 2002; 20:1879–1886.
- 108 Lindholm LH, Persson M, Alaupovic P, Carlberg B, Svensson A, Samuelsson O. Metabolic outcome during 1 year in newly detected hypertensives: results of the Antihypertensive Treatment and Lipid Profile in a North of Sweden Efficacy Evaluation (ALPINE study). *J Hypertens* 2003; 21:1563–1574.
- 109 Lindholm LH, Ibsen H, Dahlof B, Devereux RB, Beevers G, de Faire U, et al. Cardiovascular morbidity and mortality in patients with diabetes in the Losartan Intervention For Endpoint reduction in hypertension study (LIFE): a randomised trial against atenolol. *Lancet* 2002; **359**:1004–1010.
- 110 Yusuf S, Sleight P, Pogue J, Bosch J, Davies R, Dagenais G; Heart Outcomes Prevention Evaluation Study Investigators. Effects of an angiotensin-converting-enzyme inhibitor, ramipril, on cardiovascular events in high-risk patients. N Engl J Med 2000; 342:145–153.
- 111 Jansson PA. Endothelial dysfunction in insulin resistance and type 2 diabetes. J Intern Med 2007; 262:173–183.
- 112 Pechanova O, Simko F. The role of nitric oxide in the maintenance of vasoactive balance. *Physiol Res* 2007; 56(Suppl 2):S7–S16.
- 113 De Caterina R. Endothelial dysfunctions: common denominators in vascular disease. *Curr Opin Lipidol* 2000; 11:9–23.

- 114 Perticone F, Ceravolo R, Pujia A, Ventura G, Iacopino S, Scozzafava A, *et al.* Prognostic significance of endothelial dysfunction in hypertensive patients. *Circulation* 2001; **104**:191–196.
- 115 Halcox JP, Schenke WH, Zalos G, Mincemoyer R, Prasad A, Waclawiw MA, *et al.* Prognostic value of coronary vascular endothelial dysfunction. *Circulation* 2002; **106**:653–658.
- 116 Ribeiro-Oliveira A Jr, Nogueira AI, Pereira RM, Boas WW, Dos Santos RA, Simoes e Silva AC. The renin-angiotensin system and diabetes: an update. *Vasc Health Risk Manag* 2008; **4**:787–803.
- 117 Sechi LA, Griffin CA, Schambelan M. The cardiac renin-angiotensin system in STZ-induced diabetes. *Diabetes* 1994; **43**:1180–1184.
- 118 Candido R, Allen TJ, Lassila M, Cao Z, Thallas V, Cooper ME, *et al.* Irbesartan but not amlodipine suppresses diabetes-associated atherosclerosis. *Circulation* 2004; **109**:1536–1542.
- 119 Werner N, Kosiol S, Schiegl T, Ahlers P, Walenta K, Link A, et al. Circulating endothelial progenitor cells and cardiovascular outcomes. N Engl J Med 2005; 353:999–1007.
- 120 Bahlmann FH, de Groot K, Mueller O, Hertel B, Haller H, Fliser D. Stimulation of endothelial progenitor cells: a new putative therapeutic effect of angiotensin II receptor antagonists. *Hypertension* 2005; 45:526–529.
- 121 Yagil Y, Yagil C. Hypothesis: ACE2 modulates blood pressure in the mammalian organism. *Hypertension* 2003; **41**:871–873.
- 122 Watanabe T, Barker TA, Berk BC. Angiotensin II and the endothelium: diverse signals and effects. *Hypertension* 2005; **45**:163–169.
- 123 Porsti I, Bara AT, Busse R, Hecker M. Release of nitric oxide by angiotensin-(1-7) from porcine coronary endothelium: implications for a novel angiotensin receptor. *Br J Pharmacol* 1994; 111:652–654.
- 124 Brosnihan KB, Li P, Ferrario CM. Angiotensin-(1-7) dilates canine coronary arteries through kinins and nitric oxide. *Hypertension* 1996; 27(3 Pt 2):523–528.
- 125 Maia LG, Ramos MC, Fernandes L, de Carvalho MH, Campagnole-Santos MJ, Souza dos Santos RA. Angiotensin-(1-7) antagonist A-779 attenuates the potentiation of bradykinin by captopril in rats. *J Cardiovasc Pharmacol* 2004; 43:685–691.
- 126 Freeman EJ, Chisolm GM, Ferrario CM, Tallant EA. Angiotensin-(1-7) inhibits vascular smooth muscle cell growth. *Hypertension* 1996; 28:104–108.
- 127 Fraga-Silva RA, Pinheiro SV, Goncalves AC, Alenina N, Bader M, Santos RA. The antithrombotic effect of angiotensin-(1-7) involves mas-mediated NO release from platelets. *Mol Med* 2008; 14:28–35.
- 128 Pereira RM, Dos Santos RA, Teixeira MM, Leite VH, Costa LP, da Costa Dias FL, *et al.* The renin-angiotensin system in a rat model of hepatic fibrosis: evidence for a protective role of angiotensin-(1-7). *J Hepatol* 2007; **46**:674–681.
- 129 Benter IF, Yousif MH, Dhaunsi GS, Kaur J, Chappell MC, Diz DI. Angiotensin-(1-7) prevents activation of NADPH oxidase and renal vascular dysfunction in diabetic hypertensive rats. *Am J Nephrol* 2008; 28:25–33.
- 130 Kostenis E, Milligan G, Christopoulos A, Sanchez-Ferrer CF, Heringer-Walther S, Sexton PM, *et al.* G-protein-coupled receptor Mas is a physiological antagonist of the angiotensin II type 1 receptor. *Circulation* 2005; **111**:1806–1813.
- 131 Uusitupa MI, Niskanen LK, Siitonen O, Voutilainen E, Pyorala K. 5-Year incidence of atherosclerotic vascular disease in relation to general risk factors, insulin level, and abnormalities in lipoprotein composition in non-insulin-dependent diabetic and nondiabetic subjects. *Circulation* 1990; 82:27–36.

- 132 Schieffer B, Schieffer E, Hilfiker-Kleiner D, Hilfiker A, Kovanen PT, Kaartinen M, et al. Expression of angiotensin II and interleukin 6 in human coronary atherosclerotic plaques: potential implications for inflammation and plaque instability. *Circulation* 2000; 101: 1372–1378.
- 133 Gross CM, Gerbaulet S, Quensel C, Krämer J, Mittelmeier HO, Luft FC, et al. Angiotensin II type 1 receptor expression in human coronary arteries with variable degrees of atherosclerosis. Basic Res Cardiol 2002; 97:327–333.
- 134 Boring L, Gosling J, Cleary M, Charo IF. Decreased lesion formation in CCR2–/– mice reveals a role for chemokines in the initiation of atherosclerosis. *Nature* 1998; **394**:894–897.
- 135 Wassmann S, Stumpf M, Strehlow K, Schmid A, Schieffer B, Böhm M, et al. Interleukin-6 induces oxidative stress and endothelial dysfunction by overexpression of the angiotensin II type 1 receptor. *Circ Res* 2004; **94**:534–541.
- 136 Jandeleit-Dahm K, Cooper ME. Hypertension and diabetes: role of the renin-angiotensin system. *Endocrinol Metab Clin North Am* 2006; 35:469–490, vii.
- 137 Rabbani R, Topol EJ. Strategies to achieve coronary arterial plaque stabilization. *Cardiovasc Res* 1999; **41**:402–417.
- 138 Zulli A, Burrell LM, Widdop RE, Black MJ, Buxton BF, Hare DL. Immunolocalization of ACE2 and AT2 receptors in rabbit atherosclerotic plaques. *J Histochem Cytochem* 2006; **54**:147–150.
- 139 Burrell LM, Risvanis J, Kubota E, Dean RG, MacDonald PS, Lu S, *et al.* Myocardial infarction increases ACE2 expression in rat and humans. *Eur Heart J* 2005; **26**:369–375; discussion 22–24.
- 140 Calkin AC, Forbes JM, Smith CM, Lassila M, Cooper ME, Jandeleit-Dahm KA, et al. Rosiglitazone attenuates atherosclerosis in a model of insulin insufficiency independent of its metabolic effects. Arterioscler Thromb Vasc Biol 2005; 25:1903–1909.
- 141 Lewis P, Stefanovic N, Pete J, Calkin AC, Giunti S, Thallas-Bonke V, et al. Lack of the antioxidant enzyme glutathione peroxidase-1 accelerates atherosclerosis in diabetic apolipoprotein E-deficient mice. *Circulation* 2007; 115:2178–2187.
- 142 Chew P, Yuen DY, Koh P, Stefanovic N, Febbraio MA, Kola I, et al. Site-specific antiatherogenic effect of the antioxidant ebselen in the diabetic apolipoprotein E-deficient mouse. Arterioscler Thromb Vasc Biol 2009; 29:823–830.
- 143 Mancini GB, Henry GC, Macaya C, O'Neill BJ, Pucillo AL, Carere RG, et al. Angiotensin-converting enzyme inhibition with quinapril improves endothelial vasomotor dysfunction in patients with coronary artery disease. The TREND (Trial on Reversing ENdothelial Dysfunction) Study. Circulation 1996; 94:258–265.
- 144 O'Driscoll G, Green D, Maiorana A, Stanton K, Colreavy F, Taylor R. Improvement in endothelial function by angiotensin-converting enzyme inhibition in non-insulin-dependent diabetes mellitus. J Am Coll Cardiol 1999; 33:1506–1511.
- 145 Cheetham C, Collis J, O'Driscoll G, Stanton K, Taylor R, Green D. Losartan, an angiotensin type 1 receptor antagonist, improves endothelial function in non-insulin-dependent diabetes. J Am Coll Cardiol 2000; 36:1461–1466.
- 146 Hornig B, Landmesser U, Kohler C, Ahlersmann D, Spiekermann S, Christoph A, et al. Comparative effect of ace inhibition and angiotensin II type 1 receptor antagonism on bioavailability of nitric oxide in patients with coronary artery disease: role of superoxide dismutase. *Circulation* 2001; **103**:799–805.
- 147 Komers R, Simkova R, Kazdova L, Ruzickova J, Pelikanova T. Effects of ACE inhibition and AT1-receptor blockade on haemodynamic

responses to L-arginine in Type 1 diabetes. J Renin Angiotensin Aldosterone Syst 2004; 5:33–38.

- 148 Niskanen L, Hedner T, Hansson L, Lanke J, Niklason A. Reduced cardiovascular morbidity and mortality in hypertensive diabetic patients on first-line therapy with an ACE inhibitor compared with a diuretic/beta-blocker-based treatment regimen: a subanalysis of the Captopril Prevention Project. *Diabetes Care* 2001; **24**:2091–2096.
- 149 Brenner BM, Cooper ME, de Zeeuw D, Keane WF, Mitch WE, Parving HH, et al. Effects of losartan on renal and cardiovascular outcomes in patients with type 2 diabetes and nephropathy. N Engl J Med 2001; 345:861–869.
- 150 UK Prospective Diabetes Study Group. Efficacy of atenolol and captopril in reducing risk of macrovascular and microvascular complications in type 2 diabetes: UKPDS 39. Br Med J 1998; 317:713– 720.
- 151 Heart Outcomes Prevention Evaluation Study Investigators. Effects of ramipril on cardiovascular and microvascular outcomes in people with diabetes mellitus: results of the HOPE study and MICRO-HOPE substudy. *Lancet* 2000; **355**:253–259.
- 152 Tatti P, Pahor M, Byington RP, Di Mauro P, Guarisco R, Strollo G, et al. Outcome results of the Fosinopril Versus Amlodipine Cardiovascular Events Randomized Trial (FACET) in patients with hypertension and NIDDM. *Diabetes Care* 1998; 21:597–603.
- 153 Fox KM. Efficacy of perindopril in reduction of cardiovascular events among patients with stable coronary artery disease: randomised, double-blind, placebo-controlled, multicentre trial (the EUROPA study). *Lancet* 2003; **362**:782–788.
- 154 Pitt B, O'Neill B, Feldman R, Schwartz L, Mudra H, Bass T, *et al.* The QUinapril Ischemic Event Trial (QUIET): evaluation of chronic ACE inhibitor therapy in patients with ischemic heart disease and preserved left ventricular function. *Am J Cardiol* 2001; 87:1058–1063.
- 155 Friedrich EB, Teo KK, Bohm M. ACE inhibition in secondary prevention: are the results controversial? *Clin Res Cardiol* 2006; 95:61–67.
- 156 Boner G, Cooper ME, McCarroll K, Brenner BM, de Zeeuw D, Kowey PR, *et al.* Adverse effects of left ventricular hypertrophy in the reduction of endpoints in NIDDM with the angiotensin II antagonist losartan (RENAAL) study. *Diabetologia* 2005; **48**:1980–1987.
- 157 Yusuf S, Teo K, Anderson C, Pogue J, Dyal L, Copland I, et al. Telmisartan Randomised AssessmeNt Study in ACE iNtolerant subjects with cardiovascular Disease (TRANSCEND) Investigators. Effects of the angiotensin-receptor blocker telmisartan on cardiovascular events in high-risk patients intolerant to angiotensinconverting enzyme inhibitors: a randomised controlled trial. *Lancet* 2008; **372**:1174–1183.
- 158 Yanagisawa M, Kurihara H, Kimura S, Tomobe Y, Kobayashi M, Mitsui Y, *et al.* A novel potent vasoconstrictor peptide produced by vascular endothelial cells. *Nature* 1988; **332**:411–415.
- 159 Sakurai T, Yanagisawa M, Takuwa Y, Miyazaki H, Kimura S, Goto K, et al. Cloning of a cDNA encoding a non-isopeptide-selective subtype of the endothelin receptor. *Nature* 1990; **348**:732–735.
- 160 Perfetto F, Tarquini R, Tapparini L, Tarquini B. Influence of noninsulin-dependent diabetes mellitus on plasma endothelin-1 levels in patients with advanced atherosclerosis. J Diabetes Complication 1998; 12:187–192.
- 161 Kalogeropoulou K, Mortzos G, Migdalis I, Velentzas C, Mikhailidis DP, Georgiadis E, *et al.* Carotid atherosclerosis in type 2 diabetes

mellitus: potential role of endothelin-1, lipoperoxides, and prostacyclin. *Angiology* 2002; **53**:279–285.

- 162 Cunningham ME, Huribal M, Bala RJ, McMillen MA. Endothelin-1 and endothelin-4 stimulate monocyte production of cytokines. *Crit Care Med* 1997; 25:958–964.
- 163 Babaei S, Picard P, Ravandi A, Monge JC, Lee TC, Cernacek P, et al. Blockade of endothelin receptors markedly reduces atherosclerosis in LDL receptor deficient mice: role of endothelin in macrophage foam cell formation. *Cardiovasc Res* 2000; **48**:158–167.
- 164 Woods M, Wood EG, Bardswell SC, Bishop-Bailey D, Barker S, Wort SJ, et al. Role for nuclear factor-κB and signal transducer and activator of transcription 1/interferon regulatory factor-1 in cytokineinduced endothelin-1 release in human vascular smooth muscle cells. *Mol Pharmacol* 2003; 64:923–931.
- 165 Schneider MP, Boesen EI, Pollock DM. Contrasting actions of endothelin ET(A) and ET(B) receptors in cardiovascular disease. *Annu Rev Pharmacol Toxicol* 2007; 47:731–759.
- 166 King JM, Srivastava KD, Stefano GB, Bilfinger TV, Bahou WF, Magazine HI. Human monocyte adhesion is modulated by endothelin B receptor-coupled nitric oxide release. *J Immunol* 1997; 158:880–886.
- 167 Lam HC. Role of endothelin in diabetic vascular complications. Endocrine 2001; 14:277–284.
- 168 Ferri C, Laurenti O, Bellini C, Faldetta MR, Properzi G, Santucci A, et al. Circulating endothelin-1 levels in lean non-insulin-dependent diabetic patients: influence of ACE inhibition. Am J Hypertens 1995; 8:40–47.
- 169 Douglas SA, Sulpizio AC, Piercy V, Sarau HM, Ames RS, Aiyar NV, et al. Differential vasoconstrictor activity of human urotensin-II in vascular tissue isolated from the rat, mouse, dog, pig, marmoset and cynomolgus monkey. Br J Pharmacol 2000; 131:1262–1274.
- 170 Ames RS, Sarau HM, Chambers JK, Willette RN, Aiyar NV, Romanic AM, et al. Human urotensin-II is a potent vasoconstrictor and agonist for the orphan receptor GPR14. Nature 1999; 401:282–286.
- 171 Cheung BM, Leung R, Man YB, Wong LY. Plasma concentration of urotensin II is raised in hypertension. J Hypertens 2004; 22:1341–1344.
- 172 Totsune K, Takahashi K, Arihara Z, Sone M, Ito S, Murakami O. Increased plasma urotensin II levels in patients with diabetes mellitus. *Clin Sci (Lond)* 2003; **104**:1–5.
- 173 Pakala R. Role of urotensin II in atherosclerotic cardiovascular diseases. *Cardiovasc Revasc Med* 2008; **9**:166–178.
- 174 Suguro T, Watanabe T, Ban Y, Kodate S, Misaki A, Hirano T, et al. Increased human urotensin II levels are correlated with carotid atherosclerosis in essential hypertension. Am J Hypertens 2007; 20:211–217.
- 175 Wang ZJ, Shi LB, Xiong ZW, Zhang LF, Meng L, Bu DF, et al. Alteration of vascular urotensin II receptor in mice with apolipoprotein E gene knockout. *Peptides* 2006; 27:858–863.
- 176 Rakowski E, Hassan GS, Dhanak D, Ohlstein EH, Douglas SA, Giaid A. A role for urotensin II in restenosis following balloon angioplasty: use of a selective UT receptor blocker. J Mol Cell Cardiol 2005; 39:785–791.
- 177 Segain JP, Rolli-Derkinderen M, Gervois N, Raingeard de la Bletiere D, Loirand G, Pacaud P. Urotensin II is a new chemotactic factor for UT receptor-expressing monocytes. *J Immunol* 2007; 179:901–909.

- 178 Wiley SR, Schooley K, Smolak PJ, Din WS, Huang CP, Nicholl JK, et al. Identification and characterization of a new member of the TNF family that induces apoptosis. *Immunity* 1995; 3:673–682.
- 179 Marsters SA, Pitti RM, Donahue CJ, Ruppert S, Bauer KD, Ashkenazi A. Activation of apoptosis by Apo-2 ligand is independent of FADD but blocked by CrmA. *Curr Biol* 1996; 6:750–752.
- 180 Simonet WS, Lacey DL, Dunstan CR, Kelley M, Chang MS, Lüthy R, *et al.* Osteoprotegerin: a novel secreted protein involved in the regulation of bone density. *Cell* 1997; 89:309–319.
- 181 Yasuda H, Shima N, Nakagawa N, Yamaguchi K, Kinosaki M, Mochizuki S, *et al.* Osteoclast differentiation factor is a ligand for osteoprotegerin/osteoclastogenesis-inhibitory factor and is identical to TRANCE/RANKL. *Proc Natl Acad Sci U S A* 1998; 95:3597– 3602.
- 182 Emery JG, McDonnell P, Burke MB, Deen KC, Lyn S, Silverman C, et al. Osteoprotegerin is a receptor for the cytotoxic ligand TRAIL. J Biol Chem 1998; 273:14363–14367.
- 183 Secchiero P, Candido R, Corallini F, Zacchigna S, Toffoli B, Rimondi E, et al. Systemic tumor necrosis factor-related apoptosis-inducing ligand delivery shows antiatherosclerotic activity in apolipoprotein E-null diabetic mice. Circulation 2006; 114:1522–1530.
- 184 Schoppet M, Sattler AM, Schaefer JR, Hofbauer LC. Osteoprotegerin (OPG) and tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) levels in atherosclerosis. *Atherosclerosis* 2006; 184:446– 447.
- 185 Michowitz Y, Goldstein E, Roth A, Afek A, Abashidze A, Ben Gal Y, et al. The involvement of tumor necrosis factor-related apoptosisinducing ligand (TRAIL) in atherosclerosis. J Am Coll Cardiol 2005; 45:1018–1024.
- 186 Bennett BJ, Scatena M, Kirk EA, Rattazzi M, Varon RM, Averill M, et al. Osteoprotegerin inactivation accelerates advanced atherosclerotic lesion progression and calcification in older ApoE–/– mice. Arterioscler Thromb Vasc Biol 2006; 26:2117–2124.
- 187 Yano K, Tsuda E, Washida N, Kobayashi F, Goto M, Harada A, et al. Immunological characterization of circulating osteoprotegerin/osteoclastogenesis inhibitory factor: increased serum concentrations in postmenopausal women with osteoporosis. J Bone Miner Res 1999; 14:518–527.
- 188 Hofbauer LC, Schoppet M. Osteoprotegerin gene polymorphism and the risk of osteoporosis and vascular disease. J Clin Endocrinol Metab 2002; 87:4078–4079.
- 189 Knudsen ST, Foss CH, Poulsen PL, Andersen NH, Mogensen CE, Rasmussen LM. Increased plasma concentrations of osteoprotegerin in type 2 diabetic patients with microvascular complications. *Eur J Endocrinol* 2003; 149:39–42.
- 190 Hofbauer LC, Schoppet M. Osteoprotegerin: a link between osteoporosis and arterial calcification? *Lancet* 2001; 358:257–259.
- 191 Hansen TK, Tarnow L, Thiel S, Steffensen R, Stehouwer CD, Schalkwijk CG, et al. Association between mannose-binding lectin and vascular complications in type 1 diabetes. *Diabetes* 2004; 53:1570–1576.
- 192 Saraheimo M, Forsblom C, Hansen TK, Teppo AM, Fagerudd J, Pettersson-Fernholm K, *et al.* Increased levels of mannan-binding lectin in type 1 diabetic patients with incipient and overt nephropathy. *Diabetologia* 2005; **48**:198–202.
- 193 Hovind P, Hansen TK, Tarnow L, Thiel S, Steffensen R, Flybjerg A, et al. Mannose-binding lectin as a predictor of microalbuminuria in type 1 diabetes: an inception cohort study. *Diabetes* 2005; 54:1523–1527.

- 194 Hansen TK, Gall MA, Tarnow L, Thiel S, Stehouwer CD, Schalkwijk CG, et al. Mannose-binding lectin and mortality in type 2 diabetes. Arch Intern Med 2006; 166:2007–2013.
- 195 Verrier ED, Shernan SK, Taylor KM, Van de Werf F, Newman MF, Chen JC, *et al.* Terminal complement blockade with pexelizumab during coronary artery bypass graft surgery requiring cardiopulmonary bypass: a randomized trial. *JAMA* 2004; **291**:2319–2327.
- 196 Hansson L, Zanchetti A, Carruthers SG, Dahlöf B, Elmfeldt D, Julius S, et al.; HOT Study Group. Effects of intensive blood-pressure lowering and low-dose aspirin in patients with hypertension: principal results of the Hypertension Optimal Treatment (HOT) randomised trial. *Lancet* 1998; 351:1755–1762.
- 197 UK Prospective Diabetes Study Group. Tight blood pressure control and risk of macrovascular and microvascular complications in type 2 diabetes: UKPDS 38. *Br Med J* 1998; **317**:703–713.
- 198 Estacio RO, Jeffers BW, Hiatt WR, Biggerstaff SL, Gifford N, Schrier RW. The effect of nisoldipine as compared with enalapril on cardiovascular outcomes in patients with non-insulin-dependent diabetes and hypertension. *N Engl J Med* 1998; **338**:645–652.
- 199 de Galan BE, Perkovic V, Ninomiya T, Pillai A, Patel A, Cass A, et al. Lowering blood pressure reduces renal events in type 2 diabetes. J Am Soc Nephrol 2009; 20:883–892.
- 200 Yusuf S, Teo KK, Pogue J, Dyal L, Copland I, Schumacher H, et al.; ONTARGET Investigators. Telmisartan, ramipril, or both in patients at high risk for vascular events. N Engl J Med 2008; 358:1547–1559.
- 201 Collins R, Armitage J, Parish S, Sleigh P, Peto R. MRC/BHF Heart Protection Study of cholesterol-lowering with simvastatin in 5963 people with diabetes: a randomised placebo-controlled trial. *Lancet* 2003; **361**:2005–2016.
- 202 Colhoun HM, Betteridge DJ, Durrington PN, Hitman GA, Neil HA, Livingstone SJ, *et al.* Primary prevention of cardiovascular disease

with atorvastatin in type 2 diabetes in the Collaborative Atorvastatin Diabetes Study (CARDS): multicentre randomised placebocontrolled trial. *Lancet* 2004; **364**:685–696.

- 203 American Diabetes Association. Management of dyslipidemia in adults with diabetes. *Diabetes Care* 2000; **23**(Suppl 1):S57–60.
- 204 Bhatt DL. What makes platelets angry: diabetes, fibrinogen, obesity, and impaired response to antiplatelet therapy? J Am Coll Cardiol 2008; 52:1060–1061.
- 205 Jackson SP, Calkin AC. The clot thickens: oxidized lipids and thrombosis. *Nat Med* 2007; **13**:1015–1016.
- 206 Berry C, Tardif JC, Bourassa MG. Coronary heart disease in patients with diabetes: II. Recent advances in coronary revascularization. *J Am Coll Cardiol* 2007; **49**:643–656.
- 207 Keech A, Simes RJ, Barter P, Best J, Scott R, Taskinen MR, et al. Effects of long-term fenofibrate therapy on cardiovascular events in 9795 people with type 2 diabetes mellitus (the FIELD study): randomised controlled trial. *Lancet* 2005; **366**:1849–1861.
- 208 Dormandy JA, Charbonnel B, Eckland DJ, Erdmann E, Massi-Benedetti M, Moules IK, *et al.* Secondary prevention of macrovascular events in patients with type 2 diabetes in the PROactive Study (Prospective Pioglitazone Clinical Trial in Macrovascular Events): a randomised controlled trial. *Lancet* 2005; **366**:1279–1289.
- 209 Home PD, Pocock SJ, Beck-Nielsen H, Curtis PS, Gomis R, Hanefeld M, et al. Rosiglitazone evaluated for cardiovascular outcomes in oral agent combination therapy for type 2 diabetes (RECORD): a multicentre, randomised, open-label trial. Lancet 2009; 373:2125–2135.
- 210 Nissen SE, Wolski K, Topol EJ. Effect of muraglitazar on death and major adverse cardiovascular events in patients with type 2 diabetes mellitus. JAMA 2005; 294:2581–2586.