

Radiation hybrid mapping of 75 previously unreported equine microsatellite loci

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Source/description: Horse genomic DNA was digested with the restriction enzyme *Mbo*I, size selected by gel electrophoresis for fragments between 200 and 1200 bp, and ligated into the *Bam*HI site of the M13 phage vector. Clones containing a potential microsatellite were identified by screening the library with [³²P] 5' end-labelled oligo [dCA]₁₆ and oligo [dGT]₁₆ probes. The DNA was isolated from positive plaques and the inserts were sequenced using an ABI 3100 automated sequencer. Primer pairs for PCR amplification of the markers were developed using the PRIMER program (Version 0.5; M. J. Daly, S. E. Lincoln and E. S. Lander, unpublished). Sequence accession numbers, repeat motif, PCR primer pairs and product sizes based on the cloned sequences are provided in Table 1.

PCR conditions: Polymerase chain reactions for determination of microsatellite polymorphisms were performed in 15 µl volumes consisting of the following reagents: 25 ng DNA; 1X PCR buffer (Qiagen, Valencia, CA, USA); 1.5 mM MgCl₂; 25 µM each of dCTP, dGTP and dTTP; 6.25 µM dATP; 0.125 µCi [α -³²P] dATP; 0.45 U HotStar *Taq* polymerase (Qiagen) and 5 pmol of each primer. Amplification was done using a MJ Research PTC100 thermocycler (Watertown, MA, USA) under the following conditions: initial 20 min denaturation at 95 °C; 30 cycles of 94 °C for 30 s, annealing temperatures that ranged from 54–62 °C for 30 s and 72 °C for 30 s; and a final 5 min extension at 72 °C. Reaction products were electrophoresed through 7% acrylamide denaturing gels on BioRad

SequiGen GT 38 cm ×50-cm plate sequencing gel units (Hercules, CA, USA), in the presence of 1X TBE, and fragments detected using autoradiography.

The PCR for typing markers on the 5000 rad whole genome equine radiation hybrid panel comprising 92 horse × hamster hybrid cell lines¹ contained 50 ng DNA; 5 pmol of each primer; 2.0 mM MgCl₂; 0.3 U HotStar *Taq* Polymerase (Qiagen) and 10X PCR buffer which contained Tris-Cl, KCl and (NH₄)₂SO₄ (Qiagen). Markers were typed in duplicate, separated by electrophoresis on 2.5% agarose gels, and scored manually.

Allele frequencies: The DNA from 12 stallions of the Equine Genome Mapping Workshop International Reference Family,² and/or three horses (one stallion and two mares) from the parental generation of the Newmarket full-sibling reference family,³ was amplified. The number of alleles observed in this sample population ranged from 1 to 10 (Table 1).

Radiation hybrid mapping and chromosomal assignments: The PCR typing of the 75 microsatellite markers on the 5000 rad equine RH panel and data analysis was performed as described in detail by us.¹ Two point analysis with the RHMAPPY software was carried out to assign markers to map positions on the current equine radiation hybrid map¹ at LOD ≥ 12.0.¹ Chromosomal assignments and the nearest linked marker for all new equine microsatellites are provided in Table 1.

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References

- 1 Chowdhary B.P. *et al.* (2003) *Genome Res* **13**, 742–51.
- 2 Guérin G. *et al.* (1999) *Anim Genet* **30**, 341–54.
- 3 Swinburne J.E. *et al.* (2000) *Genomics* **66**, 123–34.

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Table 1 Equine microsatellite loci UMN84–UMN376.

Marker	Accession number	Repeat motif	Primer 1 (5' → 3')	Primer 2 (5' → 3')	Product size (bp)	Heterozygotes	Alleles	Chromosome (by RH mapper)	Closest Marker(s) (by RH mapper)	Distance from closest marker(s) (cR)
UMN84	AY391284	CA8 N2 CA5	ACAGTTGGATATCCGTGGGAAAC	TCGTATATGTGTGCCCTCTGC	216	0	1	ECA15	COR075	0.00
UMN89	AY391285	CA18	CGACACTTCCACAGTCACTC	TCCGTGCTTTTAAAGCAAGTTCC	149	0	1	ECA1	UCD487	17.67
UMN91	AY391286	CA19	GCAACAATAAAGATACCCAAAGCAG	GATCTTGGCCAAACAACTCGTC	153	7	5	ECAX	ARSC1	11.32
UMN93	AY391287	CA13 N2 CA8	CCAGAGTTGGAGACACCTG	GGTCCATCCACTGAAACAC	167	2*	3	ECA7	LEX045	0.00
UMN97	AY391288	CA5 N2 CA5	TGTCAGTGGCATTCAAATC	TAAACCAAGTCCCAAGTC	202	0	2	ECA18	NV077	0.00
UMN100	AY391289	CA5 N8 CA6	CTGATCGAGAGGCAATTTCTG	CGTCAGCTGAGGCAACTATG	183	5	5	ECA7	HBB	2.84
UMN102	AY391290	CA9 N4 CA9	ATGCATCTCATTGCAAGAC	GCACAAGAAATTTCCAACCTC	191	0	1	ECA15	LEX046	0.00
UMN104	AY391291	CA15	TGATTTTGTTCACCTCTATGC	CCTAGAAAGTGGAAATAAATTG	156	0	1	ECA4	HTG07	0.00
UMN106	AY391292	CA7 N CA3	AATCCTGTGAGCGAGGAGTG	TGTCTGGCATCTTTAGGG	200	0	1	ECA19	AHT052	11.88
UMN110	AY391293	CA11	TCCTTCTACCTCCATGACC	GATCTAGTACAACACTCCGTGTG	141	9	5	ECA4	PTPN12	0.00
UMN112	AY391294	CA9	TGCCAAATATTGAGCAAGG	TCTCTCCCTTTTGTG	155	0	1	ECA4	LEX033	5.23
UMN114	AY391295	CA5 N2 CA7	AACCCAAAAGCAITTTGAGAAG	TCTGCACAGTTTGTACCG	129	1	2	ECA18	SG07	19.85
UMN115	AY391296	CA12 N2 CA27	TCCTCTACACTGGCCATATC	TTTCTTACGGAGTGTCTGC	168	4	10	ECA1	MYO5A	11.09
UMN125	AY391297	CA14	TGGTCTCTGAGCATAAAG	TCCTCCCTACCTCTCACTG	143	9	5	ECA14	LEX078	0.00
UMN127	AY391298	CA10	TTATAAATCACCACTGTTACACAC	TCTTGAAGCAGGATGGGC	135	0	2	ECA26	TKY275	30.11
UMN137	AY391299	CA10	CTGCTTTTACTGCTTCACTGC	GATTTGAGTCGAGGTCTGCC	114	5	5	ECA22	HMS47	2.22
UMN139	AY391300	CA13	AGACACAGTTTGGTGGATGC	GATCAAGCACATAAGGGACAC	100	5	2	ECA21	C9	2.53
UMN143	AY391301	CA13	AGAAAGGTAAACAAGAAGCC	GACACTCTGCATCACACATG	105	3	2	ECA11	NV090	2.74
UMN147	AY391302	CA10	CAGACCTACTCCAGTCATCAGC	AAACAAGAGACTTGAAGTGGC	180	0	1	ECA21	HTG10	8.23
UMN152	AY391303	CA11	ACAGGTGTTATCCAAATTTG	AAAGTAGAGGAAGACAGGCACG	144	5	2	ECA24	COR087	0.00
UMN157	AY391304	CA11	TTCAACTGTGTGAGTGAATG	AGCTTTTGTCCAGCAGGAAC	94	5	3	ECA31	PCMT1	5.23
UMN158	AY391305	CA22	AATTGAGAGCCAAAGATGACACC	GGCACCAATTTGAGGAAGATG	146	8	8	ECA3	KCC1	4.60
UMN160	AY391306	CA7 A2 CA15	TGGAAGGATTTCCCCCAAG	GTTGCTCCAATGCCATACTC	114	0	1	ECA25	NV043	9.10
UMN164	AY391307	CA9	AAAGAGGAGGATGGCAATG	ATCTCCAGTGTGACAATCTTACC	105	0	1	ECA8	DSC2	11.43
UMN168	AY391308	CA10	CACAAAACCCCACTGAATTC	CACTACCTTCCCCTACGTTCC	149	1	2	ECA12	IGF2	20.58
UMN170	AY391309	CA7 N2 CA5	GGGTGTTAAGATCCTCTCC	CTAGGCAAACTACTGACCCCC	138	0	1	ECA20	UM011	0.00
UMN172	AY391310	CA4 N2 CA6	CCAGACACAGACCTTAGCACC	CCACAAGGAGGAGGAAAG	190	0	1	ECA4	HMS62	6.72
UMN173	AY391311	CA11	TTGGAGAAITCAAGGGTTTCC	GCCAAAGACATGGAAACAACC	132	7	5	ECA17	COR032	13.35
UMN178	AY391312	CA11	TGGCAGATAAAGCCCTTACC	GATCACATAGTGAAGGTGTTAAACG	107	1	2	ECA3	LEX007	0.00
UMN180	AY391313	CA8 N2 CA2	TGGAAAATCTCACAAAATGTC	TAATATTTCTTTTGGGTGTC	148	0	1	ECA18	CHRNA	5.34
UMN181	AY391314	CA6 N CA5	TTAAGAACCCCACTGTGTGG	AGAGAAGAGAAATCGGGGAG	149	0	1	ECA31	PCMT1	7.80
UMN182	AY391315	CA10	TCTCTCAATTCACCTCTCGG	TGAGAGAAGAGCCCAAGAAAG	132	3*	3	ECA3	KCC1	4.60
UMN183	AY391316	CA11	AGAGAACAGGAGAAGACATGCC	ACCCTGTCTCATGGAAAGCTG	106	1	2	ECA22	ADA	11.32
UMN188	AY391317	CA8 N CA2	GTTAACAAAGGATGTTTTGGGC	TGGCTTCTGCTTCTCCC	132	2	2	ECA26	EB2E8	18.15
UMN195	AY391318	CA12	CAAAAGAGTGGCCCTCGTAG	GATCTGGCCCCAGAGAAAAA	104			ECA6	MAP2	11.32
UMN196	AY391319	CA16	ATGAAAAAAGGCATACCCCC	TCTTATCAGGTTGATGGTGTGC	131	7	5	ECA1	RET	14.73

UMNe217	AY391320	CA11	CTTTGAGTTCACCAGTTCTCCC	AACCAAAAGGAACCTTTGGTGG	151	3	2	ECA13	TKY031	8.34
UMNe223	AY391321	CA10	GTCTGTCCAGACATATACCCC	AACCTGGCAAGGAAAG	112	0*	1	ECA12	TKY286	0.00
UMNe225	AY391322	CA17	GATCTCTGTGTGTTGTG	TCAATTTACAAGATGCAAAACG	154	2*	4	ECA20	UM011	0.00
UMNe227	AY391323	CA13	ATAATTTCCCTTGGCAACAC	CTGTAGACCCAAAGGAAGATGG	188	1*	2	ECA14	COR104	41.70
UMNe230	AY391324	CA11	TGGTCTAGAGTACACATACCAAC	TTTCTCTTCCACCCAG	109	0*	1	ECA5	AHT024	0.00
UMNe235	AY391325	CA17	TGAAGCAATGACAAATATCC	CACCAACAAAGTATCTGCCTG	178	2*	2	ECA17	LEX055	13.47
UMNe238	AY391326	CA11	CCAGCACCACTGTTCAAGAC	TCTAGCCATATGTTACCCTGCC	198	0*	1	ECA8	ASB14	12.33
UMNe245	AY391327	CA10	TTGCATAGGCAATCACTACG	ACAAAACCTACCCTCCG	104			ECA15	ASB19	0.00
UMNe253	AY391328	CA23	CAGAAAGATTTCTCTTTATGGC	ACTFACACTGCCTTGGAAATACC	222			ECA29	LOC91452	1.41
UMNe256	AY391329	CA14	GGCCACACACTACTTTAC	ACTTCTGGAAGACAGCTGGC	142	2*	3	ECA4	COR047	10.09
UMNe264	AY391330	CA12	GCAATGCTAGCACAGGGG	CCTCTCTCTTTCCCCCTC	101	8	3	ECA16	PDCD61P	7.26
UMNe276	AY391331	CA17	CATGCTCTCAGACTTTTCC	CACCCAGGACTCCACAG	154	2*	3	ECA22	AHT030	11.65
UMNe301	AY391332	CA5 N2 CA5	TTTTGGGTCAGGAATAAAC	TAGACAACCAATGCCAAAGG	140	4	3	ECA20	NV005	2.22
UMNe305	AY391333	CA13	ACAGTTCACAGCGGCCTC	GAGAGGAGCAACCCCTC	162	2	2	ECA13	TKY031	7.04
UMNe313	AY391334	CA15	AGAACCTGTTGAGATACGAGG	ATCAGAGTGGAGACATGGGG	179	7	5	ECA27	COR040	14.62
UMNe314	AY391335	CA10	AGGCCCTTGTATATCCAC	GAGCTGGGAGTGGAAAGGAG	153	0	1	ECA11	UCD039	0.00
UMNe318	AY391336	CA4 N2 CA7	CAGGGCTAGTCAACAGGCTC	TGGGTGTAAATGACTACCAGG	209	0	1	ECA1	COR046	4.71
UMNe319	AY391337	CA10	TTGTGCAATCTGTTGCTTC	GATCACAGTCCCTGGTACTGG	154	1	2	ECA19	HTG24	0.00
UMNe323	AY391338	CA8 CA8 CA6 CA14	GATCTGCAGGAAGCATGT	CCGCTCGGAATATTTCAATG	185	4	7	ECA2	COR037	0.00
UMNe327	AY391339	CA16	TTTTCTTCTCATTGGTGC	GAATGACAGGGCTAAGGATG	157	4	2	ECA21	C9	2.33
UMNe329	AY391340	CA19 N2 CA8	TGTTTGTGGAATGGTACG	TCTCCATCTCCACATAGGCC	104	4	3	ECA12	TKY286	8.12
UMNe330	AY391341	CA10	AAACATGGAACACAGAGGGG	TCTCCAGCGTATTTGGTTAGTG	129	0	2	ECA2	FABP3	5.55
UMNe337	AY391342	CA17	CTTTAATATGCTCGCACTTCC	TTGGGAAGTCCAGGATCTTG	215	2	3	ECA6	CUL3	0.00
UMNe350	AY391343	CA10	GCAAAATAAAGGCTCACTTGC	AGTGCTCAGGCTTATATACC	153	6	2	ECAX	COR091	0.00
UMNe351	AY391344	CA14	TGAGAAGCTTTGCCAATCC	TAGAGGCCATTTTGCATCC	150	0	1	ECA19	HTG23	2.43
UMNe353	AY391345	CA19	CCACTTTCCCTTCTCTC	AACCTTGAAGCAACCTAAGTGC	322	2*	3	ECA6	UCD465	0.00
UMNe355	AY391346	CA13	TCATGCCAATCTGAAAAGAGC	CAGAGCAAATCTTCTCACC	182	2*	2	ECA22	TKY285	12.90
UMNe357	AY391347	CA10	GATCCAGTAACAAAAGCTAATTT	TCAAATGATTTCTTACTGAGGATTC	210	1	2	ECA23	UMNe51	0.00
UMNe359	AY391348	CA10 N2 CA5 N2 CA20	GTGTGACGGGAGCAGG	TGGTGCCATCCACTGAAAC	140			ECA7	LEX045	0.00
UMNe361	AY391349	CA14	AGGGAATAACTTATGAACAGATGAA	TCITTTATGAAATTCCTCTGAACC	169	2*	3	ECA5	HMS05	5.13
UMNe362	AY391350	CA10	GAGAGAGAGATGCGCGC	GATCCAAAGGCTATCCACTC	117	2*	3	ECA25	TXN	19.48
UMNe365	AY391351	CA10	CCCTGCTCAGACTTCTGG	GAGGCTTGCATGTGGAC	201	0*	1	ECA11	PDE6G	0.00
UMNe366	AY391352	CA11	TGAACCTCAAAGCCAGGC	CATGTTCTGTAGTTTGCAGGC	174	0*	2	ECA29	TKY112	7.15
UMNe368	AY391353	CA15	TCCAGAAAAGCAACACTCAC	TGTCAGTTCCAGATTTTGGCC	292	0*	1	ECA31	VIP	0.00
UMNe372	AY391354	CA11	ACGTTACAGGTGGGGAGATG	CTTGGCTTCCAGGAAAACAC	127			ECA10	CASP8A2	3.67
UMNe373	AY391355	CA17	AGGCCACTCAAGCCACAG	CACCTCTCTCCAAATGAG	265	9	4	ECA15	ASB19	0.00
UMNe374	AY391356	CA16	AATTTATATGTTTCCATGCC	TAAGCACTGCGTTAATGTTCTG	194	2*	3	ECA2	SMARCA5	31.61
UMNe375	AY391357	CA6 N2 CA12	GGAGCCAGTTCAAAAGATTTCC	ATTGAGTGGTGTGTTGGCC	130	2*	2	ECAX	FACL4	3.98
UMNe376	AY391358	CA21	ACGCATATAAAGTTTGAGAAATCC	CAGACCTACTGAATGGAAATACACC	155	2*	5	ECAX	COL4A5	3.15

Repeat motif, PCR primers and product sizes, polymorphism and radiation hybrid map location (chromosomal assignment along with closest located marker and distance in cR) for these new microsatellite markers were obtained as described in the accompanying text. An asterisk (*) by the number of heterozygous individuals indicates that these markers were only typed on the Animal Health Trust reference family parents.